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(71) Applicant (for all designated States except US): DANISCO A/S [DK/DK]; Langebrogade 1, P.O. Box 17, DK-1001 Copenhagen K (DK).

(72) Inventor; and

(75) Inventor/Applicant (for US only): POULSEN, Peter [DK/DK]; Danisco a/s, Langebrogade 1, P.O. Box 17, DK-1001 Copenhagen K (DK).

(74) Agents: MASCHIO, Antonio et al.; D Young & Co., 21 New Fetter Lane, London EC4A 1DA (GB).

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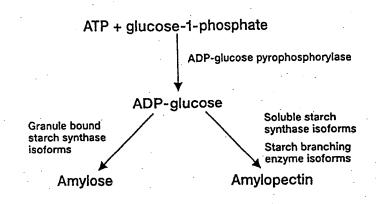
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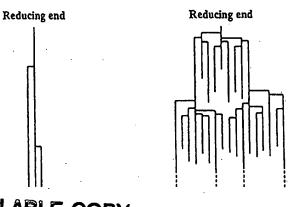
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(54) Title: SENSE INTRON INHIBITION OF STARCH BRANCHING ENZYME EXPRESSION

(57) Abstract

A method of inhibiting gene expression is described. The method, which affects enzymatic activity in a plant, comprises expressing in a plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for a class A SBE intron in a sense orientation; and wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron.





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SENSE INTRON INHIBITION OF STARCH BRANCHING ENZYME EXPRESSION

The present invention relates to a method of inhibiting gene expression, particularly inhibiting gene expression in a plant. The present invention also relates to a nucleotide sequence useful in the method. In addition, the present invention relates to a promoter that is useful for expressing the nucleotide sequence.

Starch is one of the main storage carbohydrates in plants, especially higher plants. The structure of starch consists of amylose and amylopectin. Amylose consists essentially of straight chains of α -1-4-linked glycosyl residues. Amylopectin comprises chains of α -1-4-linked glycosyl residues with some α -1-6 branches. The branched nature of amylopectin is accomplished by the action of *inter alia* an enzyme commonly known as the starch branching enzyme ("SBE"). SBE catalyses the formation of branch points in the amylopectin molecule by adding α -1,4 glucans through α -1,6-glucosidic branching linkages. The biosynthesis of amylose and amylopectin is schematically shown in Figure 1, whereas the α -1-4-links and the α -1-6 links are shown in Figure 2.

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In Potato, it is known that two classes of SBE exist. In our copending international patent applications PCT/EP96/03052 and PCT/EP96/03053, class B potato SBE and a gene encoding it are discussed. In international patent application WO96/34968, class A potato SBE and a cDNA encoding it are disclosed.

It is known that starch is an important raw material. Starch is widely used in the food, paper, and chemical industries. However, a large fraction of the starches used in these industrial applications are post-harvest modified by chemical, physical or enzymatic methods in order to obtain starches with certain required functional properties.

Within the past few years it has become desirable to make genetically modified plants which could be capable of producing modified starches which could be the same as the post-harvest modified starches. It is also known that it may be possible to prepare such genetically modified plants by expression of antisense nucleotide coding sequences. In

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this regard, June Bourque provides a detailed summary of antisense strategies for the genetic manipulations in plants (Bourque 1995 Plant Science 105 pp 125-149).

WO96/34968 discusses the use of antisense sequences complementary to sequences which encode class A and class B potato SBE to downregulate SBE expression in potato plants.

The sequences used are complementary to SBE coding sequences.

Whilst it is known that enzymatic activity can be affected by expression of particular nucleotide sequences (for example see the teachings of Finnegan and McElroy [1994] Biotechnology 12 883-888; and Matzke and Matzke [1995] TIG 11 1-3) there is still a need for a method that can more reliably and/or more efficiently and/or more specifically affect enzymatic activity.

According to a first aspect of the present invention there is provided a method of affecting enzymatic activity in a plant (or a cell, a tissue or an organ thereof) comprising expressing in the plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence partially or completely codes (is) an intron of the potato class A SBE gene in a sense orientation, optionally together with a nucleotide sequence which codes, partially or completely, for an intron of a class B starch branching enzyme in a sense or antisense orientation; and wherein the nucleotide sequence does not contain a sequence that is a sense exon sequence normally associated with the intron.

According to a second aspect of the present invention there is provided a method of affecting enzymatic activity in a starch producing organism (or a cell, a tissue or an organ thereof) comprising expressing in the starch producing organism (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for an intron of the potato class A SBE gene in a sense orientation optionally together with a nucleotide sequence which codes, partially or completely, for an intron of a class B starch branching enzyme in a sense or antisense orientation; wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron; and wherein starch branching enzyme activity is

affected and/or the levels of amylopectin are affected and/or the composition of starch is changed.

Preferably, the class A SBE gene sense intron construct is used in combination with a potato class B SBE gene sense intron construct as defined in PCT/EP96/03053. However, it may also be used independently thereof, to target class A SBE alone, or in combination with other transgenes such as other sense and/or antisense transgenes, for example antisense intron transgenes such as from SBE genes, to further manipulate starch quality in potato plants.

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According to a third aspect of the present invention there is provided a sequence comprising the nucleotide sequence shown as SEQ. ID. No. 38 or a variant, derivative or homologue thereof.

According to a fourth aspect of the present invention there is provided a promoter comprising the sequence shown as SEQ.I.D. No. 14 or a variant, derivative or homologue thereof.

According to a fifth aspect of the present invention there is provided a construct capable of comprising or expressing the present invention.

According to a sixth aspect of the present invention there is provided a vector comprising or expressing the present invention.

According to a seventh aspect of the present invention there is provided a cell, tissue or organ comprising or expressing the present invention.

According to an eighth aspect of the present invention there is provided a transgenic starch producing organism comprising or expressing the present invention. According to a ninth aspect of the present invention there is provided a starch obtained from the present invention.

A key advantage of the present invention is that it provides a method for preparing modified starches that is not dependent on the need for post-harvest modification of starches. Thus the method of the present invention obviates the need for the use of hazardous chemicals that are normally used in the post-harvest modification of starches.

In addition, the present invention provides *inter alia* genetically modified plants which are capable of producing modified and/or novel and/or improved starches whose properties would satisfy various industrial requirements.

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Thus, the present invention provides a method of preparing tailor-made starches in plants which could replace the post-harvest modified starches.

Also, the present invention provides a method that enables modified starches to be prepared by a method that can have a more beneficial effect on the environment than the known post-harvest modification methods which are dependent on the use of hazardous chemicals and large quantities of energy.

An other key advantage of the present invention is that it provides a method that may more reliably and/or more efficiently and/or more specifically affect enzymatic activity when compared to the known methods of affecting enzymatic activity. With regard to this advantage of the present invention it is to be noted that there is some degree of homology between coding regions of SBEs. However, there is little or no homology with the intron sequences of SBEs. Thus, sense intron expression provides a mechanism to affect selectively the expression of a particular SBE. This advantageous aspect could be used, for example, to reduce or eliminate a particular SBE enzyme and replace that enzyme with another enzyme which can be another branching enzyme or even a recombinant version of the affected enzyme or even a hybrid enzyme which could for example comprise part of a SBE enzyme from one source and at least a part of another SBE enzyme from another source. This particular feature of the present invention is

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covered by the combination aspect of the present invention which is discussed in more detail later.

Thus the present invention provides a mechanism for selectively affecting SBE activity.

This is in contrast to the prior art methods which are dependent on the use of for example antisense exon expression whereby it would not be possible to introduce new SBE activity without affecting that activity as well.

In the context of the present invention, class B SBE is synonymous with SBE I: class A SBE is synonymous with SBE II. Class A SBE is as defined in WO96/34968, incorporated herein by reference. Preferably, the antisense intron construct used comprises intron 1 of class A SBE, which is 2.0 kb in length and is located starting at residue 45 of the coding sequence of class A SBE. The boundaries of the intron may be calculated by searching for consensus intron boundary sequences, and are shown in attached figure 11. The sequence of the intron is set forth in SEQ. ID. No. 38. Class B SBE is substantially as defined in the sequences given herein and in PCT/EP96/03053.

Preferably with the first aspect of the present invention starch branching enzyme activity is affected and/or wherein the levels of amylopectin are affected and/or the composition of starch is changed.

Preferably with the first or second aspect of the present invention the nucleotide sequence does not contain a sequence that is sense to an exon sequence.

25 Preferably with the fourth aspect of the present invention the promoter is in combination with a gene of interest ("GOI").

Preferably the enzymatic activity is reduced or eliminated.

Preferably the nucleotide sequence codes for at least substantially all of at least one intron in a sense orientation.

Preferably the nucleotide sequence codes, partially or completely, for two or more introns and wherein each intron is in a sense orientation.

5 Preferably the nucleotide sequence comprises at least 350 nucleotides (e.g. 350 bp), more preferably at least 500 nucleotides (e.g. 500 bp).

Preferably the nucleotide sequence comprises the sequence shown as SEQ. ID. No. 38, or a fragment thereof.

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Preferably the nucleotide sequence is expressed by a promoter having a sequence shown as SEQ. I.D. No. 14 or a variant, derivative or homologue thereof.

Preferably the transgenic starch producing organism is a plant.

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A preferred aspect of the present invention therefore relates to a method of affecting enzymatic activity in a plant (or a cell, a tissue or an organ thereof) comprising expressing in the plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for a class A SBE intron in a sense orientation; wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron; and wherein starch branching enzyme activity is affected and/or the levels of amylopectin are affected and/or the composition of starch is changed.

A more preferred aspect of the present invention therefore relates to a method of affecting enzymatic activity in a plant (or a cell, a tissue or an organ thereof) comprising expressing in the plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for an intron in a sense orientation; wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron; wherein starch branching enzyme activity is affected and/or the levels of amylopectin are affected and/or the composition of

starch is changed; and wherein the nucleotide sequence comprises the sequence shown as SEQ. ID. No. 38, or fragments thereof.

The term "nucleotide" in relation to the present invention includes DNA and RNA.

Preferably it means DNA, more preferably DNA prepared by use of recombinant DNA techniques.

The term "intron" is used in its normal sense as meaning a segment of nucleotides, usually DNA, that does not encode part or all of an expressed protein or enzyme.

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The term "exon" is used in its normal sense as meaning a segment of nucleotides, usually DNA, encoding part or all of an expressed protein or enzyme.

Thus, the term "intron" refers to gene regions that are transcribed into RNA molecules, but which are spliced out of the RNA before the RNA is translated into a protein. In contrast, the term "exon" refers to gene regions that are transcribed into RNA and subsequently translated into proteins.

The terms "variant" or "homologue" or "fragment" in relation to the nucleotide sequence of the present invention include any substitution of, variation of, modification of, replacement of, deletion of or addition of one (or more) nucleic acid from or to the respective nucleotide sequence providing the resultant nucleotide sequence can affect enzyme activity in a plant, or cell or tissue thereof, preferably wherein the resultant nucleotide sequence has at least the same effect as the sequence shown in SEQ. ID. No. 38. In particular, the term "homologue" covers homology with respect to similarity of structure and/or similarity of function providing the resultant nucleotide sequence has the ability to affect enzymatic activity in accordance with the present invention. With respect to sequence homology (i.e. similarity), preferably there is more than 80% homology, more preferably at least 85% homology, more preferably at least 90% homology, even more preferably at least 95% homology, more preferably at least 98% homology. The above terms are also synonymous with allelic variations of the sequences.

Likewise, the terms "variant" or "homologue" or "fragment" in relation to the promoter of the present invention include any substitution of, variation of, modification of, replacement of, deletion of or addition of one (or more) nucleic acid from or to the respective promoter sequence providing the resultant promoter sequence allows expression of a GOI, preferably wherein the resultant promoter sequence has at least the same effect as SEQ.I.D. No. 14. In particular, the term "homologue" covers homology with respect to similarity of structure and/or similarity of function providing the resultant promoter sequence has the ability to allow for expression of a GOI, such as a nucleotide sequence according to the present invention. With respect to sequence homology (i.e. similarity), preferably there is more than 80% homology, more preferably at least 85% homology, more preferably at least 90% homology, even more preferably at least 95% homology, more preferably at least 98% homology. The above terms are also synonymous with allelic variations of the sequences.

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The intron sequence of the present invention can be any one or all of the intron sequences of the present invention, including partial sequences thereof, provided that if partial sense sequences are used the partial sequences affect enzymatic activity. Suitable examples of partial sequences include sequences that are shorter than any one of the full sense sequences shown as SEQ. ID. No. 38 but which comprise nucleotides that are adjacent the respective exon or exons.

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With regard to the second aspect of the present invention (i.e. specifically affecting SBE activity), the nucleotide sequences of the present invention may comprise one or more sense or antisense exon sequences of the class A or class B SBE gene (but not sense exon sequences naturally associated with the intron sequence), including complete or partial sequences thereof, providing the nucleotide sequences can affect SBE activity, preferably wherein the nucleotide sequences reduce or eliminate SBE activity. Preferably, the nucleotide sequence of the second aspect of the present invention does not comprise sense exon sequences.

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The term "vector" includes an expression vector and a transformation vector. The term "expression vector" means a construct capable of *in vivo* or *in vitro* expression. The term "transformation vector" means a construct capable of being transferred from one species to another - such as from an *E.Coli* plasmid to a fungus or a plant cell, or from an *Agrobacterium* to a plant cell.

The term "construct" - which is synonymous with terms such as "conjugate", "cassette" and "hybrid" - in relation to the sense nucleotide sequence aspect of the present invention includes the nucleotide sequence according to the present invention directly or indirectly attached to a promoter. An example of an indirect attachment is the provision of a suitable spacer group such as an intron sequence, such as the *Sh1*-intron or the ADH intron, intermediate the promoter and the nucleotide sequence of the present invention. The same is true for the term "fused" in relation to the present invention which includes direct or indirect attachment. The terms do not cover the natural combination of the wild type SBE gene when associated with the wild type SBE gene promoter in their natural environment.

The construct may even contain or express a marker which allows for the selection of the genetic construct in, for example, a plant cell into which it has been transferred. Various markers exist which may be used in, for example, plants - such as mannose. Other examples of markers include those that provide for antibiotic resistance - e.g. resistance to G418, hygromycin, bleomycin, kanamycin and gentamycin.

The construct of the present invention preferably comprises a promoter. The term "promoter" is used in the normal sense of the art, e.g. an RNA polymerase binding site in the Jacob-Monod theory of gene expression. Examples of suitable promoters are those that can direct efficient expression of the nucleotide sequence of the present invention and/or in a specific type of cell. Some examples of tissue specific promoters are disclosed in WO 92/11375.

The promoter could additionally include conserved regions such as a Pribnow Box or a TATA box. The promoters may even contain other sequences to affect (such as to maintain, enhance, decrease) the levels of expression of the nucleotide sequence of the present invention. Suitable examples of such sequences include the *Sh1*-intron or an ADH intron. Other sequences include inducible elements - such as temperature, chemical, light or stress inducible elements. Also, suitable elements to enhance transcription or translation may be present. An example of the latter element is the TMV 5' leader sequence (see Sleat Gene 217 [1987] 217-225; and Dawson Plant Mol. Biol. 23 [1993] 97).

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As mentioned, the construct and/or the vector of the present invention may include a transcriptional initiation region which may provide for regulated or constitutive expression. Any suitable promoter may be used for the transcriptional initiation region, such as a tissue specific promoter. In one aspect, preferably the promoter is the patatin promoter or the E35S promoter. In another aspect, preferably the promoter is the SBE promoter.

If, for example, the organism is a plant then the promoter can be one that affects expression of the nucleotide sequence in any one or more of seed, tuber, stem, sprout, root and leaf tissues, preferably tuber. By way of example, the promoter for the nucleotide sequence of the present invention can be the α -Amy 1 promoter (otherwise known as the Amy 1 promoter, the Amy 637 promoter or the α -Amy 637 promoter) as described in our co-pending UK patent application No. 9421292.5 filed 21 October 1994. Alternatively, the promoter for the nucleotide sequence of the present invention can be the α -Amy 3 promoter (otherwise known as the Amy 3 promoter, the Amy 351 promoter or the α -Amy 351 promoter) as described in our co-pending UK patent application No. 9421286.7 filed 21 October 1994.

The present invention also encompasses the use of a promoter to express a nucleotide sequence according to the present invention, wherein a part of the promoter is inactivated but wherein the promoter can still function as a promoter. Partial inactivation of a

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promoter in some instances is advantageous. In particular, with the Amy 351 promoter mentioned earlier it is possible to inactivate a part of it so that the partially inactivated promoter expresses the nucleotide sequence of the present invention in a more specific manner such as in just one specific tissue type or organ. The term "inactivated" means partial inactivation in the sense that the expression pattern of the promoter is modified but wherein the partially inactivated promoter still functions as a promoter. However, as mentioned above, the modified promoter is capable of expressing a gene coding for the enzyme of the present invention in at least one (but not all) specific tissue of the original promoter. Examples of partial inactivation include altering the folding pattern of the promoter sequence, or binding species to parts of the nucleotide sequence, so that a part of the nucleotide sequence is not recognised by, for example, RNA polymerase. Another, and preferable, way of partially inactivating the promoter is to truncate it to form fragments thereof. Another way would be to mutate at least a part of the sequence so that the RNA polymerase can not bind to that part or another part. Another modification is to mutate the binding sites for regulatory proteins for example the CreA protein known from filamentous fungi to exert carbon catabolite repression, and thus abolish the catabolite repression of the native promoter.

The construct and/or the vector of the present invention may include a transcriptional termination region.

The nucleotide according to the present invention can be expressed in combination (but not necessarily at the same time) with an additional construct. Thus the present invention also provides a combination of constructs comprising a first construct comprising the nucleotide sequence according to the present invention operatively linked to a first promoter; and a second construct comprising a GOI operatively linked to a second promoter (which need not be the same as the first promoter). With this aspect of the present invention the combination of constructs may be present in the same vector, plasmid, cells, tissue, organ or organism. This aspect of the present invention also covers methods of expressing the same, preferably in specific cells or tissues, such as expression in just a specific cell or tissue, of an organism, typically a plant. With this aspect of the

present invention the second construct does not cover the natural combination of the gene coding for an enzyme ordinarily associated with the wild type gene promoter when they are both in their natural environment.

An example of a suitable combination would be a first construct comprising the nucleotide sequence of the present invention and a promoter, such as the promoter of the present invention, and a second construct comprising a promoter, such as the promoter of the present invention, and a GOI wherein the GOI codes for another starch branching enzyme either in sense or antisense orientation.

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The above comments relating to the term "construct" for the sense nucleotide aspect of the present invention are equally applicable to the term "construct" for the promoter aspect of the present invention. In this regard, the term includes the promoter according to the present invention directly or indirectly attached to a GOI.

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The term "GOI" with reference to the promoter aspect of the present invention or the combination aspect of the present invention means any gene of interest, which need not necessarily code for a protein or an enzyme - as is explained later. A GOI can be any nucleotide sequence that is either foreign or natural to the organism in question, for example a plant.

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Typical examples of a GOI include genes encoding for other proteins or enzymes that modify metabolic and catabolic processes. The GOI may code for an agent for introducing or increasing pathogen resistance.

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The GOI may even be an antisense construct for modifying the expression of natural transcripts present in the relevant tissues. An example of such a GOI is the nucleotide sequence according to the present invention.

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The GOI may even code for a protein that is non-natural to the host organism - e.g. a plant. The GOI may code for a compound that is of benefit to animals or humans. For example, the GOI could code for a pharmaceutically active protein or enzyme such as any one of the therapeutic compounds insulin, interferon, human serum albumin, human growth factor and blood clotting factors. The GOI may even code for a protein giving additional nutritional value to a food or feed or crop. Typical examples include plant proteins that can inhibit the formation of anti-nutritive factors and plant proteins that have a more desirable amino acid composition (e.g. a higher lysine content than a non-transgenic plant). The GOI may even code for an enzyme that can be used in food processing such as xylanases and α -galactosidase. The GOI can be a gene encoding for any one of a pest toxin, an antisense transcript such as that for α -amylase, a protease or a glucanase. Alternatively, the GOI can be a nucleotide sequence according to the present invention.

- The GOI can be the nucleotide sequence coding for the arabinofuranosidase enzyme which is the subject of our co-pending UK patent application 9505479.7. The GOI can be the nucleotide sequence coding for the glucanase enzyme which is the subject of our co-pending UK patent application 9505475.5. The GOI can be the nucleotide sequence coding for the α-amylase enzyme which is the subject of our co-pending UK patent application 9413439.2. The GOI can be the nucleotide sequence coding for the α-amylase enzyme which is the subject of our co-pending UK patent application 9421290.9. The GOI can be any of the nucleotide sequences coding for the α-glucan lyase enzyme which are described in our co-pending PCT patent application PCT/EP94/03397.
- In one aspect the GOI can even be a nucleotide sequence according to the present invention but when operatively linked to a different promoter.

The GOI could include a sequence that codes for one or more of a xylanase, an arabinase, an acetyl esterase, a rhamnogalacturonase, a glucanase, a pectinase, a branching enzyme or another carbohydrate modifying enzyme or proteinase. Alternatively, the GOI may be a sequence that is antisense to any of those sequences.

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As mentioned above, the present invention provides a mechanism for selectively affecting a particular enzymatic activity.

In an important application of the present invention it is now possible to reduce or eliminate expression of a genomic nucleotide sequence coding for a genomic protein or enzyme by expressing a sense intron construct for that particular genomic protein or enzyme and (e.g. at the same time) expressing a recombinant version of that enzyme or protein - in other words the GOI is a recombinant nucleotide sequence coding for the genomic enzyme or protein. This application allows expression of desired recombinant enzymes and proteins in the absence of (or reduced levels of) respective genomic enzymes and proteins. Thus the desired recombinant enzymes and proteins can be easily separated and purified from the host organism. This particular aspect of the present invention is very advantageous over the prior art methods which, for example, rely on the use of antisense exon expression which methods also affect expression of the recombinant enzyme.

Thus, a further aspect of the present invention relates to a method of expressing a recombinant protein or enzyme in a host organism comprising expressing a nucleotide sequence coding for the recombinant protein or enzyme; and expressing a further nucleotide sequence wherein the further nucleotide sequence codes, partially or completely, for an intron in a sense-orientation; wherein the intron is an intron normally associated with the genomic gene encoding a protein or an enzyme corresponding to the recombinant protein or enzyme; and wherein the further nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron. Additional aspects cover the combination of those nucleotide sequences including their incorporation in constructs, vectors, cells, tissues and transgenic organisms.

Therefore the present invention also relates to a combination of nucleotide sequences comprising a first nucleotide sequence coding for a recombinant enzyme; and a second nucleotide sequence which corresponds to an intron in a sense orientation; wherein the intron is an intron that is associated with a genomic gene encoding the enzyme

corresponding to the recombinant enzyme; and wherein the second nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron.

- The GOI may even code for one or more introns but in an antisense orientation, such as any one or more of the antisense intron sequences presented in the attached sequence listings. For example, the present invention also covers the expression of for example a sense intron (e.g. SEQ.I.D.No. 38) in combination with for example an antisense intron which preferably is not complementary to the sense intron sequence (e.g. SEQ.I.D.No. 10).
 - The terms "cell", "tissue" and "organ" include cell, tissue and organ per se and when within an organism.
- The term "organism" in relation to the present invention includes any organism that could comprise the nucleotide sequence according to the present invention and/or wherein the nucleotide sequence according to the present invention can be expressed when present in the organism. Preferably the organism is a starch producing organism such as any one of a plant, algae, fungi, yeast and bacteria, as well as cell lines thereof. Preferably the organism is a plant.

The term "starch producing organism" includes any organism that can biosynthesise starch. Preferably, the starch producing organism is a plant.

- The term "plant" as used herein includes any suitable angiosperm, gymnosperm, monocotyledon and dicotyledon. Typical examples of suitable plants include vegetables such as potatoes; cereals such as wheat, maize, and barley; fruit; trees; flowers; and other plant crops. Preferably, the term means "potato".
- 30 The term "transgenic organism" in relation to the present invention includes any organism that comprises the nucleotide sequence according to the present invention and/or products

obtained therefrom, and/or wherein the nucleotide sequence according to the present invention can be expressed within the organism. Preferably the nucleotide sequence of the present invention is incorporated in the genome of the organism. Preferably the transgenic organism is a plant, more preferably a potato.

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To prepare the host organism one can use prokaryotic or eukaryotic organisms. Examples of suitable prokaryotic hosts include *E. coli* and *Bacillus subtilis*. Teachings on the transformation of prokaryotic hosts is well documented in the art, for example see Sambrook *et al* (Sambrook *et al*. in Molecular Cloning: A Laboratory Manual, 2nd edition, 1989, Cold Spring Harbor Laboratory Press).

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Even though the enzyme according to the present invention and the nucleotide sequence coding for same are not disclosed in EP-B-0470145 and CA-A-2006454, those two documents do provide some useful background commentary on the types of techniques that may be employed to prepare transgenic plants according to the present invention. Some of these background teachings are now included in the following commentary.

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The basic principle in the construction of genetically modified plants is to insert genetic information in the plant genome so as to obtain a stable maintenance of the inserted genetic material.

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Several techniques exist for inserting the genetic information, the two main principles being direct introduction of the genetic information and introduction of the genetic information by use of a vector system. A review of the general techniques may be found in articles by Potrykus (Annu Rev Plant Physiol Plant Mol Biol [1991] 42:205-225) and Christou (Agro-Food-Industry Hi-Tech March/April 1994 17-27).

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Thus, in one aspect, the present invention relates to a vector system which carries a nucleotide sequence or construct according to the present invention and which is capable of introducing the nucleotide sequence or construct into the genome of an organism, such as a plant.

The vector system may comprise one vector, but it can comprise two vectors. In the case of two vectors, the vector system is normally referred to as a binary vector system. Binary vector systems are described in further detail in Gynheung An et al. (1980), Binary Vectors, Plant Molecular Biology Manual A3, 1-19.

One extensively employed system for transformation of plant cells with a given promoter or nucleotide sequence or construct is based on the use of a Ti plasmid from Agrobacterium tumefaciens or a Ri plasmid from Agrobacterium rhizogenes An et al. (1986), Plant Physiol. 81, 301-305 and Butcher D.N. et al. (1980), Tissue Culture Methods for Plant Pathologists, eds.: D.S. Ingrams and J.P. Helgeson, 203-208. Several different Ti and Ri plasmids have been constructed which are suitable for the construction of the plant or plant cell constructs described above. A non-limiting example of such a Ti plasmid is pGV3850.

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The nucleotide sequence or construct of the present invention should preferably be inserted into the Ti-plasmid between the terminal sequences of the T-DNA or adjacent a T-DNA sequence so as to avoid disruption of the sequences immediately surrounding the T-DNA borders, as at least one of these regions appears to be essential for insertion of modified T-DNA into the plant genome.

As will be understood from the above explanation, if the organism is a plant the vector system of the present invention is preferably one which contains the sequences necessary to infect the plant (e.g. the *vir* region) and at least one border part of a T-DNA sequence, the border part being located on the same vector as the genetic construct.

Furthermore, the vector system is preferably an Agrobacterium tumefaciens Ti-plasmid or an Agrobacterium rhizogenes Ri-plasmid or a derivative thereof. As these plasmids are well-known and widely employed in the construction of transgenic plants, many vector systems exist which are based on these plasmids or derivatives thereof.

In the construction of a transgenic plant the nucleotide sequence or construct of the present invention may be first constructed in a microorganism in which the vector can replicate and which is easy to manipulate before insertion into the plant. An example of a useful microorganism is *E. coli*, but other microorganisms having the above properties may be used. When a vector of a vector system as defined above has been constructed in *E. coli*, it is transferred, if necessary, into a suitable *Agrobacterium* strain, e.g. *Agrobacterium tumefaciens*. The Ti-plasmid harbouring the nucleotide sequence or construct of the present invention is thus preferably transferred into a suitable *Agrobacterium* strain, e.g. *A. tumefaciens*, so as to obtain an *Agrobacterium* cell harbouring the promoter or nucleotide sequence or construct of the present invention, which DNA is subsequently transferred into the plant cell to be modified.

If, for example, for the transformation the Ti- or Ri-plasmid of the plant cells is used, at least the right boundary and often however the right and the left boundary of the Ti- and Ri-plasmid T-DNA, as flanking areas of the introduced genes, can be connected. The use of T-DNA for the transformation of plant cells has been intensively studied and is described in EP-A-120516; Hoekema, in: The Binary Plant Vector System Offset-drukkerij Kanters B.B., Alblasserdam, 1985, Chapter V; Fraley, et al., Crit. Rev. Plant Sci., 4:1-46; and An et al., EMBO J. (1985) 4:277-284.

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Direct infection of plant tissues by Agrobacterium is a simple technique which has been widely employed and which is described in Butcher D.N. et al. (1980), Tissue Culture Methods for Plant Pathologists, eds.: D.S. Ingrams and J.P. Helgeson, 203-208. For further teachings on this topic see Potrykus (Annu Rev Plant Physiol Plant Mol Biol [1991] 42:205-225) and Christou (Agro-Food-Industry Hi-Tech March/April 1994 17-27). With this technique, infection of a plant may be performed in or on a certain part or tissue of the plant, i.e. on a part of a leaf, a root, a stem or another part of the plant.

Typically, with direct infection of plant tissues by Agrobacterium carrying the GOI (such as the nucleotide sequence according to the present invention) and, optionally, a promoter, a plant to be infected is wounded, e.g. by cutting the plant with a razor blade

or puncturing the plant with a needle or rubbing the plant with an abrasive. The wound is then inoculated with the *Agrobacterium*. The inoculated plant or plant part is then grown on a suitable culture medium and allowed to develop into mature plants.

- When plant cells are constructed, these cells may be grown and maintained in accordance with well-known tissue culturing methods such as by culturing the cells in a suitable culture medium supplied with the necessary growth factors such as amino acids, plant hormones, vitamins, etc.
- Regeneration of the transformed cells into genetically modified plants may be accomplished using known methods for the regeneration of plants from cell or tissue cultures, for example by selecting transformed shoots using an antibiotic and by subculturing the shoots on a medium containing the appropriate nutrients, plant hormones, etc.

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Further teachings on plant transformation may be found in EP-A-0449375.

As reported in CA-A-2006454, a large amount of cloning vectors are available which contain a replication system in *E. coli* and a marker which allows a selection of the transformed cells. The vectors contain for example pBR 322, pUC series, M13 mp series, pACYC 184 etc. In this way, the nucleotide or construct of the present invention can be introduced into a suitable restriction position in the vector. The contained plasmid is then used for the transformation in *E.coli*. The *E.coli* cells are cultivated in a suitable nutrient medium and then harvested and lysed. The plasmid is then recovered. As a method of analysis there is generally used sequence analysis, restriction analysis, electrophoresis and further biochemical-molecular biological methods. After each manipulation, the used DNA sequence can be restricted and connected with the next DNA sequence. Each sequence can be cloned in the same or different plasmid.

30 After the introduction of the nucleotide sequence or construct according to the present invention in the plants the presence and/or insertion of further DNA sequences may be

necessary - such as to create combination systems as outlined above (e.g. an organism comprising a combination of constructs).

The above commentary for the transformation of prokaryotic organisms and plants with the nucleotide sequence of the present invention is equally applicable for the transformation of those organisms with the promoter of the present invention.

In summation, the present invention relates to affecting enzyme activity by expressing sense intron sequences.

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Also, the present invention relates to a promoter useful for the expression of those sense intron sequences.

The following samples have been deposited in accordance with the Budapest Treaty at the recognised depositary The National Collections of Industrial and Marine Bacteria Limited (NCIMB) at 23 St Machar Drive, Aberdeen, Scotland, AB2 1RY, United Kingdom, on 13 July 1995:

NCIMB 40754 (which refers to pBEA 11 as described herein);

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NCIMB 40751 (which refers to λ -SBE 3.2 as described herein), and

NCIMB 40752 (which refers to λ -SBE 3.4 as described herein).

A highly preferred embodiment of the present invention therefore relates to a method of affecting enzymatic activity in a plant (or a cell, a tissue or an organ thereof) comprising expressing in the plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for an intron in a sense orientation; wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron; wherein starch branching enzyme activity is affected and/or the levels of amylopectin are affected and/or the composition of

starch is changed; and wherein the intron nucleotide sequence is the sequence of intron 1 of class A SBE as set forth in SEQ. ID. No. 38, or any other intron of class A SBE, including fragments thereof, and including combinations of class A sense intron sequences and class B sense or antisense intron sequences. The sequence of introns of class A SBE other than intron 1 may be obtained by sequencing of, for example, potato class A SBE genomic DNA, isolatable by hybridisation screening of a genomic DNA library with class A SBE cDNA obtainable according to WO96/34968 according to methods well known in the art and set forth, for example, in Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor, 1989.

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The present invention will now be described only by way of example, in which reference is made to the following attached Figures:

Figure 1, which is a schematic representation of the biosynthesis of amylose and amylopectin;

Figure 2, which is a diagrammatic representation of the α -1-4-links and the α -1-6 links of amylopectin;

Figure 3, which is a diagrammatic representation of the exon-intron structure of a genomic SBE clone;

Figure 4, which is a plasmid map of pPATA1, which is 3936 bp in size;

25 Figure 5, which is a plasmid map of pABE7, which is 5106 bp in size;

Figure 6, which is a plasmid map of pVictorIV Man, which is 7080 bp in size;

Figure 7, which is a plasmid map of pBEA11, which is 9.54 kb in size;

Figure 8, which shows the full genomic nucleotide sequence for SBE including the promoter, exons and introns;

Figure 9, which is a plasmid map of pVictor5a, which is 9.12 kb in size;

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Figure 10, which is a plasmid map of pBEP2, which is 10.32 kb in size;

Figure 11, which shows the positioning of intron 1 in the class A and class B SBE genes;

10 Figure 12, which shows the sequence of intron 1 of the potato class A SBE;

Figure 13, which shows pSS15; and

Figure 14, which shows pSS16.

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Figures 1 and 2 were referred to above in the introductory description concerning starch in general. As mentioned, Figure 3 is a diagrammatic representation of the exon-intron structure of a genomic SBE clone, the sequence of which is shown in Figure 8. This clone, which has about 11.5 k base pairs, comprises 14 exons and 13 introns. The introns are numbered in increasing order from the 5' end to the 3' end and correspond to SEQ.I.D.No.s 1-13, respectively. Their respective antisense intron sequences are shown as SEQ.I.D.No.s 15-27.

In more detail, Figures 3 and 8 present information on the 11468 base pairs of a potato SBE gene. The 5' region from nucleotides 1 to 2082 contain the promoter region of the SBE gene. A TATA box candidate at nucleotide 2048 to 2051 is boxed. The homology between a potato SBE cDNA clone (Poulsen & Kreiberg (1993) Plant Physiol 102: 1053-1054) and the exon DNAs begin at 2083 bp and end at 9666 bp. The homology between the cDNA and the exon DNA is indicated by nucleotides in upper case letters, while the translated amino acid sequences are shown in the single letter code below the exon DNA. Intron sequences are indicated by lower case letters.

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Figure 7 is a plasmid map of pBEA7, which is 9.54 k base pairs in size. Plasmid pBEA 11 comprises the first intron sequence of the potato SBE gene. This first intron sequence, which has 1177 base pairs, is shown in Figure 3 and lies between the first exon and the second exon.

These experiments and aspects of the present invention are now discussed in more detail.

10 EXPERIMENTAL PROTOCOL

ISOLATION, SUBCLONING IN PLASMIDS, AND SEQUENCING OF GENOMIC SBE CLONES

Various clones containing the potato SBE gene are isolated from a Desiree potato genomic library (Clontech Laboratories Inc., Palo Alto CA, USA) using radioactively labelled potato SBE cDNA (Poulsen & Kreiberg (1993) Plant Physiol. 102:1053-1054) as probe. The fragments of the isolated λ-phages containing SBE DNA (λSBE 3.2 - NCIMB 40751 - and λSBE-3.4 - NCIMB 40752) are identified by Southern analysis and then subcloned into pBluescript II vectors (Clontech Laboratories Inc., Palo Alto CA, USA). λSBE 3.2 contains a 15 kb potato DNA insert and λSBE-3.4 contains a 13 kb potato DNA insert. The resultant plasmids are called pGB3, pGB11, pGB15, pGB16 and pGB25 (see discussion below). The respective inserts are then sequenced using the Pharmacia Autoread Sequencing Kit (Pharmacia, Uppsala) and a A.L.F. DNA sequencer (Pharmacia, Uppsala).

In total, a stretch of 11.5 kb of the SBE gene is sequenced. The sequence is deduced from the above-mentioned plasmids, wherein: pGB25 contains the sequences from 1 bp to 836 bp, pGB15 contains the sequences from 735 bp to 2580 bp, pGB16 contains the sequences from 2580 bp to 5093 bp, pGB11 contains the sequences from 3348 bp to 7975 bp, and pGB3 contains the sequences from 7533 bp to 11468 bp.

In more detail, pGB3 is constructed by insertion of a 4 kb *EcoRI* fragment isolated from λ SBE 3.2 into the *EcoRI* site of pBluescript II SK (+). pGB11 is constructed by

insertion of a 4.7 kb XhoI fragment isolated from λSBE 3.4 into the XhoI site of pBluescript II SK (+). pGB15 is constructed by insertion of a 1.7 kb SpeI fragment isolated from λSBE 3.4 into the SpeI site of pBluescript II SK (+). pGB16 is constructed by insertion of a 2.5 kb SpeI fragment isolated from λSBE 3.4 into the SpeI site of pBluescript II SK (+). For the construction of pGB25 a PCR fragment is produced with the primers

5' GGA ATT CCA GTC GCA GTC TAC ATT AC 3' (SEQ. ID. No. 30)

and

5' CGG GAT CCA GAG GCA TTA AGA TTT CTG G 3'

(SEQ. ID. No. 31)

and $\lambda SBE 3.4$ as a template.

The PCR fragment is digested with BamHI and EcoRI, and inserted in pBluescript II SK (+) digested with the same restriction enzymes.

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CONSTRUCTION OF PLASMID pBEA11

The SBE intron 1 is amplified by PCR using the oligonucleotides

5' CGG GAT CCA AAG AAA TTC TCG AGG TTA CAT GG 3'

(SEQ. ID. No. 32)

20 and

5' CGG GAT CCG GGG TAA TTT TTA CTA ATT TCA TG 3"-

(SEO. ID. No. 33)

and the λSBE 3.4 phage containing the SBE gene as template.

The PCR product is digested with BamHI and inserted in a sense orientation in the BamHI site of plasmid pPATA1 (described in WO 94/24292) between the patatin promoter and the 35S terminator. This construction, pABE7, is digested with KpnI, and the 2.4 kb "patatin promoter-SBE intron 1-35S terminator" KpnI fragment is isolated and inserted in the KpnI site of the plant transformation vector pVictorIV Man yielding plasmid pBEA11.

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CONSTRUCTION OF PLASMID pSS15.

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The 2122 bp intron 1 sequence of the potato SBEII gene (see SEQ. ID. No. 38) is amplified by PCR from a genomic SBEII subclone using the primers 5' - CGG GAT CCC GTA TGT CTC ACT GTG TTT GTG GC - 3' (SEQ. ID. No. 34) and 5' - CGG GAT CCC CCT ACA TAC ATA TAT CAG ATT AG - 3' (SEQ. ID. No. 35). The PCR product is digested with BamHI and inserted in sense orientation after a patatin promoter in the BamHI site of a plant transformation vector in which the NPTII gene is used as selectable marker (see figure 13).

CONSTRUCTION OF PLASMID pSS16.

The 2122 bp intron 1 sequence of the potato SBEII gene (SEQ. ID. No. 38) is amplified by PCR from a genomic SBEII subclone using the primers 5' - CGG GAT CCC GTA TGT CTC ACT GTG TTT GTG GC - 3' (SEQ. ID. No. 34) and 5' - CGG GAT CCC CCT ACA TAC ATA TAT CAG ATT AG - 3' (SEQ. ID. No. 35). The PCR product is digested with BamHI and inserted in sense orientation after a patatin promoter in the BamHI site of a plant transformation vector in which the manA gene is used as selectable marker (see figure 14).

PRODUCTION OF TRANSGENIC POTATO PLANTS

Axenic stock cultures

Shoot cultures of *Solanum tuberosum* 'Bintje' and 'Dianella' are maintained on a substrate (LS) of a formula according to Linsmaier, E.U. and Skoog, F. (1965), Physiol. Plant. 18: 100-127, in addition containing 2 μ M silver thiosulphate at 25°C and 16 h light/8 h dark.

The cultures are subcultured after approximately 40 days. Leaves are then cut off the shoots and cut into nodal segments (approximately 0.8 cm) each containing one node.

Inoculation of potato tissues

Shoots from approximately 40 days old shoot cultures (height approximately 5-6 cms) are cut into internodal segments (approximately 0.8 cm). The segments are placed into liquid LS-substrate containing the transformed Agrobacterium tumefaciens containing the binary vector of interest. The Agrobacterium are grown overnight in YMB-substrate

(di-potassium hydrogen phosphate, trihydrate (0.66 g/l); magnesium sulphate, heptahydrate (0.20 g/l); sodium chloride (0.10 g/l); mannitol (10.0 g/l); and yeast extract (0.40 g/l)) containing appropriate antibiotics (corresponding to the resistance gene of the *Agrobacterium* strain) to an optical density at 660 nm (OD-660) of approximately 0.8, centrifuged and resuspended in the LS-substrate to an OD-660 of 0.5.

The segments are left in the suspension of *Agrobacterium* for 30 minutes and then the excess of bacteria are removed by blotting the segments on sterile filter paper.

Co-cultivation

The shoot-segments are co-cultured with bacteria for 48 hours directly on LS-substrate containing agar (8.0 g/l), 2,4-dichlorophenoxyacetic acid (2.0 mg/l) and transzeatin (0.5 mg/l). The substrate and also the explants are covered with sterile filter papers, and the petri dishes are placed at 25°C and 16 h light/8 dark.

"Washing" procedure

After the 48 h on the co-cultivation substrate the segments are transferred to containers containing liquid LS-substrate containing 800 mg/l carbenicillin. The containers are gently shaken and by this procedure the major part of the *Agrobacterium* is either washed off the segments and/or killed.

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Selection

After the washing procedure the segments are transferred to plates containing the LS-substrate, agar (8 g/l), trans-zeatin (1-5 mg/l), gibberellic acid (0.1 mg/l), carbenicillin (800 mg/l), and kanamycin sulphate (50-100 mg/l) or phosphinotricin (1-5 mg/l) or mannose (5 g/l) depending on the vector construction used. The segments are sub-cultured to fresh substrate each 3-4 weeks. In 3 to 4 weeks, shoots develop from the segments and the formation of new shoots continued for 3-4 months.

Rooting of regenerated shoots

The regenerated shoots are transferred to rooting substrate composed of LS-substrate, agar (8 g/l) and carbenicillin (800 mg/l). The transgenic genotype of the

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regenerated shoot are verified by testing the rooting ability on the above mentioned substrates containing kanamycin sulphate (200 mg/l), by performing NPTII assays (Radke, S. E. et al, Theor. Appl. Genet. (1988), 75: 685-694) or by performing PCR analysis according to Wang *et al* (1993, NAR 21 pp 4153-4154). Plants which are not positive in any of these assays are discarded or used as controls. Alternatively, the transgenic plants could be verified by performing a GUS assay on the co-introduced β -glucuronidase gene according to Hodal, L. *et al.* (Pl. Sci. (1992), 87: 115-122).

Transfer to soil

The newly rooted plants (height approx. 2-3 cms) are transplanted from rooting substrate to soil and placed in a growth chamber (21°C, 16 hour light 200-400uE/m²/sec). When the plants are well established they are transferred to the greenhouse, where they are grown until tubers had developed and the upper part of the plants are senescing.

15 Harvesting

The potatoes are harvested after about 3 months and then analysed.

BRANCHING ENZYME ANALYSIS

The SBE expression in the transgenic potato lines are measured using the SBE assays described by Blennow and Johansson (Phytochemistry (1991) 30:437-444) and by standard Western procedures using antibodies directed against class A and class B potato SBE.

STARCH ANALYSIS

Starch is isolated from potato tubers and analysed for the amylose:amylopectin ratio (Hovenkamp-Hermelink et al. (1988) Potato Research 31:241-246). In addition, the chain length distribution of amylopectin is determined by analysis of isoamylase digested starch on a Dionex HPAEC. The number of reducing ends in isoamylase digested starch is determined by the method described by N. Nelson (1944) J. Biol.Chem. 153:375-380.

The results revealed that there is a reduction in the level of synthesis of SBE and/or the level of activity of SBE and/or the composition of starch SBE in the transgenic plants.

5 CONSTRUCTION OF SBE PROMOTER CONSTRUCT

An SBE promoter fragment is amplified from λ -SBE 3.4 using primers:

5° CCA TCG ATA CTT TAA GTG ATT TGA TGG C 3'

(SEQ. ID. No. 36)

and

5' CGG GAT CCT GTT CTG ATT CTT GAT TTC C 3'.

(SEQ. ID. No. 37)

The PCR product is digested with ClaI and BamHI. The resultant 1.2 kb fragment is then inserted in pVictor5a (see Figure 9) linearised with ClaI and BgIII yielding pBEP2 (see Figure 10).

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STARCH BRANCHING ENZYME MEASUREMENTS OF POTATO TUBERS

Potatoes from potato plants transformed with pBEA11 are cut in small pieces and homogenised in extraction buffer (50 mM Tris-HCl pH 7.5, Sodium-dithionite (0.1 g/l), and 2 mM DTT) using a Ultra-Turax homogenizer; 1 g of Dowex xl. is added pr. 10 g of tuber. The crude homogenate is filtered through a miracloth filter and centrifuged at 4°C for 10 minutes at 24.700 g. The supernatant is used for starch branching enzyme assays. The starch branching enzyme assays are carried out at 25 °C in a volume of 400 μl composed of 0.1 M Na citrate buffer pH 7.0, 0.75 mg/ml amylose, 5 mg/ml bovine serum albumin and the potato extract. At 0, 15 30 and 60 minutes aliquouts of 50 μl are removed from the reaction into 20 μl 3 N HCl. 1 ml of iodine solution is added and the decrease in absorbance at 620 nm is measured with an ELISA spectrophotometer.

The starch branching enzyme (SBE) levels in tuber extracts are measured from 24 transgenic Dianella potato plants transformed with plasmid pBEA11, pSS15 and pSS16.

The results show that the BEA11, SS15 and SS16 transgenic lines produce tubers which have class B and class A SBE levels, respectively, that are only 10 % to 15 % of the SBE levels found in non transformed Dianella plants.

In a further experiment, plasmids pSS15 and pBEA11 are cotransfected into potato plants, as described above. In the cotransfectants, when analysed as set forth above, simultaneous reduction of class A and class B SBE levels are observed.

5 SUMMATION

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The above-mentioned examples relate to the isolation and sequencing of a gene for potato SBE. The examples further demonstrate that it is possible to prepare SBE intron constructs. These SBE intron constructs can be introduced into plants, such as potato plants. After introduction, a reduction in the level of synthesis of SBE and/or the level of activity of SBE and/or the composition of starch in plants can be achieved.

Without wishing to be bound by theory it is believed that the expressed sense intron nucleotide sequence according to the present invention affects enzymatic activity via co-suppression and/or trans-activation. Reviews of these mechanisms has been published by Finnegan and McElroy (1994 Biotechnology 12 pp 883 - 887) and Matzke and Matzke (1995 TIG 11 No. 1 pp 1 - 3). By these mechanisms, it is believed that the sense introns of the present invention reduce the level of plant enzyme activity (in particular SBE activity), which in turn for SBE activity is believed to influence the amylose:amylopectin ratio and thus the branching pattern of amylopectin.

Thus, the present invention provides a method wherein it is possible to manipulate the starch composition in plants, or tissues or cells thereof, such as potato tubers, by reducing the level of SBE activity by using sense intron sequences.

The simultaneous reduction or elimination of class A and class B SBE sequences from the doubly transformed potato plants, moreover, offers the possibility to transform such plants with different SBE genes at will, thus allowing the manipulation of branching in starch according to the desired result.

In summation the present invention therefore relates to the surprising use of SBE class A sense intron sequences in a method to affect class A SBE activity in plants.

Other modifications of the present invention will be apparent to those skilled in the art without departing from the scope of the present invention.

The following pages present a number of sequence listings which have been consecutively numbered from SEQ.I.D. No. 1 - SEQ.I.D. No. 38. In brief, SEQ.I.D. No. 1 - SEQ.I.D. No. 13 represent sense intron sequences (genomic DNA); SEQ.I.D.

No. 14 represents the SBE promoter sequence (genomic sequence); SEQ.I.D. No. 15 - SEQ.I.D. No. 27 represent antisense intron sequences; and SEQ. I.D. No. 28 represents the sequence complementary to the SBE promoter sequence - i.e. the SBE promoter sequence in antisense orientation. The full genomic nucleotide sequence for SBE including the promoter, exons and introns is shown as SEQ. I.D. No. 29 (see Figures 3 and 8 which highlight particular gene features). SEQ. ID. No. 30 to 37 show primers used in the methods set forth above. SEQ. ID. No. 38 represents the nucleotide sequence of intron 1 of the class A potato SBE gene.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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- (i) APPLICANT:
 - (A) NAME: DANISCO A/S
 - (B) STREET: LANGEBROGADE 1
 - (C) CITY: COPENHAGEN K
- 10 (E) COUNTRY: DENMARK
 - (F) POSTAL CODE (ZIP): DK-1001
 - (ii) TITLE OF INVENTION: INHIBITION OF GENE EXPRESSION
- 15 (iii) NUMBER OF SEQUENCES: 38
 - (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
- 20 (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)
 - (2) INFORMATION FOR SEQ ID NO: 1:

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- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1165 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
- 30 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO

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(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

5	GTAATTTTTA	CTAATTTCAT	GTTAATTTCA	ATTATTTTA	GCCTTTGCAT	TTCATTTTCC	60
	AATATATCTG	GATCATCTCC	TTAGTTTTTT	ATTTTATTTT	TTATAATATC	AAATATGGAA	120
	GAAAAATGAC	ACTTGTAGAG	CCATATGTAA	GTATCATGTG	ACAAATTTGC	AAGGTGGTTG	180
10	AGTGTATAAA	ATTCAAAAAT	TGAGAGATGG	AGGGGGGTG	GGGGAAGACA	ATATTTAGAA	240
٠	AGAGTGTTCT	AGGAGGTTAT	GGAGGACACG	GATGAGGGGT	AGAAGGTTAG	TTAGGTATTT	300
15	GAGTGTTGTC	TGGCTTATCC	TTTCATACTA	GTAGTCGTGG	AATTATTTGG	GTAGTTTCTT	360
	GTTTTGTTAT	TTGATCTTTG	TTATTCTATT	TTCTGTTTCT	TGTACTTCGA	TTATTGTATT	420
••	ATATATCTTG	TCGTAGTTAT	TGTTCCTCGG	TAAGAATGCT	CTAGCATGCT	TCCTTTAGTG	480
20	TTTTATCATG	CCTTCTTTAT	ATTCGCGTTG	CTTTGAAATG	CTTTTACTTT	AGCCGAGGGT	540
	CTATTAGAAA	CAATCTCTCT	ATCTCGTAAG	GTAGGGGTAA	AGTCCTCACC	ACACTCCACT	600
25	TGTGGGATTA	CATTGTGTTT	GTTGTTGTAA	ATCAATTATG	TATACATAAT	AAGTGGATTT	660
	TTTACAACAC	AAATACATGG	TCAAGGGCAA	AGTTCTGAAC	ACATAAAGGG	TTCATTATAT	720
	GTCCAGGGAT	' ATGATAAAA	TTGTTTCTT	GTGAAAGTTA	TATAAGATTI	GTTATGGCTT	780
30	TTGCTGGAAA	CATAATAAGI	TATAATGCTO	AGATAGCTAC	TGAAGTTTG	TTTTTCTAGC	840
	CTTTTAAATG	TACCAATAAT	AGATTCCGT	A TCGAACGAGI	: ATGTTTTGAT	TACCTGGTCA	900
35	TGATGTTTCT	T ATTTTTAC	\ TTTTTTTGG	r gttgaactgo	AATTGAAAA	r GTTGTATCCT	960
				m cmamccacc	ר ייכאכאאפריי	האמרכריים האמרכריים	1020

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	CCAATAATTT CTATGAATTC AAATTCAGTT TATGGCTACC AGTCAGTCCA GAAATTAGGA	1080
	TATGCTGCAT ATACTTGTTC AATTATACTG TAAAATTTCT TAAGTTCTCA AGATATCCAT	1140
5	GTAACCTCGA GAATTTCTTT GACAG	1165
	(2) INFORMATION FOR SEQ ID NO: 2:	
	(i) SEQUENCE CHARACTERISTICS:	
0	(A) LENGTH: 317 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(B) Torollog1. Illieuz	
15	(ii) MOLECULE TYPE: DNA (genomic)	
	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
20	(17) 12:11 52:152: 115	
20 .		
		-
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:	
25		
	GTATGTTTGA TAATTTATAT GGTTGCATGG ATAGTATATA AATAGTTGGA AAACTTCTGG	60
•	ACTGGTGCTC ATGGCATATT TGATCTGTGC ACCGTGTGGA GATGTCAAAC ATGTGTTACT	120
30	TCGTTCCGCC AATTTATAAT ACCTTAACTT GGGAAAGACA GCTCTTTACT CCTGTGGGCA	180
•	TTTGTTATTT GAATTACAAT CTTTATGAGC ATGGTGTTTT CACATTATCA ACTTCTTTCA	240
	TGTGGTATAT AACAGTTTTT AGCTCCGTTA ATACCTTTCT TCTTTTTGAT ATAAACTAAC	300
35	TGTGGTGCAT TGCTTGC	317

(2) INFORMATION FOR SEQ ID NO: 3:

	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 504 base pairs	
	(B) TYPE: nucleic acid	
5	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	•
10	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: NO	
15		•
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:	
	GTAACAGCCA AAAGTTGTGC TTTAGGCAGT TTGACCTTAT TTTGGAAGAT GAATTGTTTA	60
20		
	TACCTACTTT GACTTTGCTA GAGAATTTTG CATACCGGGG AGTAAGTAGT GGCTCCATTT	120
		180
	AGGTGGCACC TGGCCATTTT TTTGATCTTT TAAAAAGCTG TTTGATTGGG TCTTCAAAAA	100
25	AGTAGACAAG GTTTTTGGAG AAGTGACACA CCCCCGGAGT GTCAGTGGCA AAGCAAAGAT	240
25	AGTAGACAAG GIIIIIGGAG AAGIGACACA CCCCCGGAGI GICAGIGGGI	
	TTTCACTAAG GAGATTCAAA ATATAAAAAA AGTATAGACA TAAAGAAGCT GAGGGGATTC	300
	AACATGTACT ATACAAGCAT CAAATATAGT CTTAAAGCAA TTTTGTAGAA ATAAAGAAAG	360
30		
	TCTTCCTTCT GTTGCTTCAC AATTTCCTTC TATTATCATG AGTTACTCTT TCTGTTCGAA	420
•		
٠	ATAGCTTCCT TAATATTAAA TTCATGATAC TTTTGTTGAG ATTTAGCAGT TTTTTCTTGT	480
		504
35	GTAAACTGCT CTCTTTTTT GCAG	504
	(A) THEODMARION FOR SEC. ID NO. 4	
	(2) INFORMATION FOR SEQ ID NO: 4:	

	(i) SEQUENCE CHARACTERISTICS:			
	(A) LENGTH: 146 base pairs			
••	(B) TYPE: nucleic acid			•
	(C) STRANDEDNESS: single			
5	(D) TOPOLOGY: linear			
	(ii) MOLECULE TYPE: DNA (genomic)		·	
	(iii) HYPOTHETICAL: NO			•
10	,			
	(iv) ANTI-SENSE: NO		,	
	(14) 2011 501152 110	•	ě	
			•	•
		•		
15				
13	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:	:		,
	(A2)			
	GTAGGTCCTC GTCTACTACA AAATAGTAGT TTCCATCATC	ATAACAGATT	TTCCTATTAA	60
20	AGCATGATGT TGCAGCATCA TTGGCTTTCT TACATGTTCT	AATTGCTATT	AAGGTTATGC	120
	TTCTAATTAA CTCATCCACA ATGCAG			146
	(2) INFORMATION FOR SEQ ID NO: 5:		•	
25				
	(i) SEQUENCE CHARACTERISTICS:			
	(A) LENGTH: 218 base pairs			
	(B) TYPE: nucleic acid			
	(C) STRANDEDNESS: single			i
30	(D) TOPOLOGY: linear	•		
			•	
•	(ii) MOLECULE TYPE: DNA (genomic)			
	(iii) HYPOTHETICAL: NO			
35				

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

	•	
5	GTTTTGTTAT TCATACCTTG AAGCTGAATT TTGAACACCA TCATCACAGG CATTTCGATT	60
	CATGTTCTTA CTAGTCTTGT TATGTAAGAC ATTTTGAAAT GCAAAAGTTA AAATAATTGT	120
	GTCTTTACTA ATTTGGACTT GATCCCATAC TCTTTCCCTT AACAAATGA GTCAATTCTA	180
10	TAAGTGCTTG AGAACTTACT ACTTCAGCAA TTAAACAG	218
	(2) INFORMATION FOR SEQ ID NO: 6:	
15	(i) SEQUENCE CHARACTERISTICS:	•
	(A) LENGTH: 198 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
20		
	(ii) MOLECULE TYPE: DNA (genomic)	
	(iii) HYPOTHETICAL: NO	
25	(iv) ANTI-SENSE: NO	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:	
	GTATTTTAAA TTTATTTCTA CAACTAAATA ATTCTCAGAA CAATTGTTAG ATAGAATCCA	60
35	AATATATACG TCCTGAAAGT ATAAAAGTAC TTATTTTCGC CATGGGCCTT CAGAATATTG	120
	GTAGCCGCTG AATATCATGA TAAGTTATTT ATCCAGTGAC ATTTTTATGT TCACTCCTAT	180
	TATGTCTGCT GGATACAG	198

	(2) INFORMATION FOR SEQ ID NO: 7:
	(i) SEQUENCE CHARACTERISTICS:
5	(A) LENGTH: 208 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
	(D) TOPOLOGY: linear
10	(ii) MOLECULE TYPE: DNA (genomic)
	(iii) HYPOTHETICAL: NO
•	(iv) ANTI-SENSE: NO
15	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:
20	
	GTTTGTCTGT TTCTATTGCA TTTTAAGGTT CATATAGGTT AGCCACGGAA AATCTCACTC 60
	TTTGTGAGGT AACCAGGGTT CTGATGGATT ATTCAATTTT CTCGTTTATC ATTTGTTTAT 120
25	TCTTTTCATG CATTGTGTTT CTTTTTCAAT ATCCCTCTTA TTTGGAGGTA ATTTTTCTCA 180
	TCTATTCACT TTTAGCTTCT AACCACAG 208
30	(2) INFORMATION FOR SEQ ID NO: 8:
50	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 293 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
35	(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

10	GTATGTCTTA	CATCTTTAGA	TATTTTGTGA	TAATTACAAT	TAGTTTGGCT	TACTTGAACA	60
	AGATTCATTC	CTCAAAATGA	CCTGAACTGT	TGAACATCAA	AGGGGTTGAA	ACATAGAGGA	120
1.5	AAACAACATG	ATGAATGTTT	CCATTGTCTA	GGGATTTCTA	TTATGTTGCT	GAGAACAAAT	180
15	GTCATCTTAA	AAAAAACATT	GTTTACTTTT	TTGTAGTATA	GAAGATTACT	GTATAGAGTT	240
	TGCAAGTGTG	TCTGTTTTGG	AGTAATTGTG	AAATGTTTGA	TGAACTTGTA	CAG	293

20 (2) INFORMATION FOR SEQ ID NO: 9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 376 base pairs
 - (B) TYPE: nucleic acid
- 25 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
- 30 (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: NO

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

PCT/IB98/00295

	39	
	GTTCAAGTAT TTTGAATCGC AGCTTGTTAA ATAATCTAGT AATTTTTAGA TTGCTTACTT	60
	GGAAGTCTAC TTGGTTCTGG GGATGATAGC TCATTTCATC TTGTTCTACT TATTTTCCAA	120
5	CCGAATTTCT GATTTTGTT TCGAGATCCA AGTATTAGAT TCATTTACAC TTATTACCGC	180
	CTCATTTCTA CCACTAAGGC CTTGATGAGC AGCTTAAGTT GATTCTTTGA AGCTATAGTT	240
10	TCAGGCTACC AATCCACAGC CTGCTATATT TGTTGGATAC TTACCTTTTC TTTACAATGA	300
10	AGTGATACTA ATTGAAATGG TCTAAATCTG ATATCTATAT TTCTCCGTCT TTCCTCCCCC	360
•	TCATGATGAA ATGCAG	376
15	(2) INFORMATION FOR SEQ ID NO: 10:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 172 base pairs	·
20	(B) TYPE: nucleic acid (C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	٠.
	(ii) MOLECULE TYPE: DNA (genomic)	
25	(iii) HYPOTHETICAL: NO	. •
	(iv) ANTI-SENSE: NO	
30		
•	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:	
	GTAAAATCAT CTAAAGTTGA AAGTGTTGGG TTTATGAAGT GCTTTAATTC TATCCAAGGA	60
35	CAAGTAGAAA CCTTTTTACC TTCCATTTCT TGATGATGGA TTTCATATTA TTTAATCCAA	120
	TO COMPACT THE COMPACT THE AG	172

	(2) INFORMATION FOR SEQ ID NO: 11:			
	(i) SEQUENCE CHARACTERISTICS:			
5	(A) LENGTH: 145 base pairs	•		
-	(B) TYPE: nucleic acid			
	(C) STRANDEDNESS: single			٠.
	(D) TOPOLOGY: linear			
10	(ii) MOLECULE TYPE: DNA (genomic)			
	(iii) HYPOTHETICAL: NO		•	
	(iv) ANTI-SENSE: NO			٠.
15		٠.		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1	1:	•	
20	GTATATATGT TTTACTTATC CATGAAATTA TTGCTCTGCT	TGTTTTTAAT	GTACTGAACA	60
	AGTTTTATGG AGAAGTAACT GAAACAAATC ATTTTCACAT	TGTCTAATTT	AACTCTTTTT	120
25	TCTGATCCTC GCATGACGAA AACAG	_	•	145
	(2) INFORMATION FOR SEQ ID NO: 12:			
	(i) SEQUENCE CHARACTERISTICS:	. •		
30	(A) LENGTH: 242 base pairs		*	
30	(B) TYPE: nucleic acid			
•	(C) STRANDEDNESS: single			
	(D) TOPOLOGY: linear			
35	(ii) MOLECULE TYPE: DNA (genomic)			

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

	GTAAGGATTT	GCTTGAATAA	CTTTTGATAA	TAAGATAACA	GATGTAGGGT	ACAGTTCTCT	60
10	CACCAAAAAG	AACTGTAATT	GTCTCATCCA	TCTTTAGTTG	TATAAGATAT	CCGACTGTCT	120
-	GAGTTCGGAA	GTGTTTGAGC	CTCCTGCCCT	CCCCCTGCGT	TGTTTAGCTA	ATTCAAAAAG	180
	GAGAAAACTG	TTTATTGATG	ATCTTTGTCT	TCATGCTGAC	ATACAATCTG	TTCTCATGAC	240
15	AG						242

(2) INFORMATION FOR SEQ ID NO: 13:

20 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 797 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

30 (iv) ANTI-SENSE: NO

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

	GCGATAGAAG	TTAACTATTG	ATTACCGCCA	CAATCGCCAG	TTAAGTCCTC	TGAACTACTA	120
	ATTTGAAAGG	TAGGAATAGC	CGTAATAAGG	TCTACTTTTG	GCATCTTACT	GTTACAAAAC	180
5	AAAAGGATGC	CAAAAAAATT	CTTCTCTATC	CTCTTTTTCC	CTAAACCAGT	GCATGTAGCT	240
	TGCACCTGCA	TAAACTTAGG	TAAATGATCA	AAAATGAAGT	TGATGGGAAC	TTAAAACCGC	300
	CCTGAAGTAA	AGCTAGGAAT	AGTCATATAA	TGTCCACCTT	TGGTGTCTGC	GCTAACATCA	360
10	ACAACAACAT	ACCTCGTGTA	GTCCCACAAA	GTGGTTTCAG	GGGGAGGGTA	GAGTGTATGC	420
	AAAACTTACT	CCTATCTCAG	AGGTAGAGAG	GATTTTTCA	ATAGACCCTT	GGCTCAAGAA	480
15	AAAAAGTCCA	AAAAGAAGTA	ACAGAAGTGA	AAGCAACATG	TGTAGCTAAA	GCGACCCAAC	540
	TTGTTTGGGA	CTGAAGTAGT	TGTTGTTGTT	GAAACAGTGC	ATGTAGATGA	ACACATGTCA	600
20	GAAAATGGAC	AACACAGTTA	TTTTGTGCA	GTCAAAAAAA	TGTACTACTA	TTTCTTTGTG	660
	CAGCTTTATG	TATAGAAAAG	TTAAATAACI	T AATGAATTT	GCTAGCAGAA	AAATAGCTTG	720
	GAGAGAAATT	TTTTATATT	AACTAAGCTA	ACTATATTCA	A TCTTTCTTT	TGCTTCTTCT	780
25	TCTCCTTGTT	TGTGAAG					797

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

30

(A) LENGTH: 2169 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

35

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

	ATCATGGCCA	ATTACTGGTT	CAAATGCATT	ACTTCCTTTC	AGATTCTTTC	GAGTTCTCAT	60
0	GACCGGTCCT	ACTACAGACG	ATACTAACCC	GTGGAACTGT	TGCATCTGCT	TCTTAGAACT	120
	CTATGGCTAT	TTTCGTTAGC	TTGGCGTCGG	TTTGAACATA	GTTTTTGTTT	TCAAACTCTT	180
	CATTTACAGT	CAAAATGTTG	TATGGTTTTT	GTTTTCCTCA	ATGATGTTTA	CAGTGTTGTG	240
	TTGTCATCTG	TACTTTTGCC	TATTACTTGT	TTTGAGTTAC	ATGTTAAAAA	AGTGTTTATT	300
	TTGCCATATT	TTGTTCTCTT	ATTATTATTA	TCATACATAC	ATTATTACAA	GGAAAAGACA	360
20	AGTACACAGA	TCTTAACGTT	TATGTTCAAT	CAACTTTTGG	AGGCATTGAC	AGGTACCACA	420
	AATTTTGAGT	TTATGATTAA	GTTCAATCTT	AGAATATGAA	TTTAACATCT	ATTATAGATG	480
25	CATAAAAATA	GCTAATGATA	GAACATTGAC	AȚTTGGCAGA	GCTTAGGGTA	TGGTATATCC	540
25	AACGTTAATT	TAGTAATTT	TGTTACGTAC	GTATATGAAA	TATTGAATTA	ATCACATGAA	600
•	CGGTGGATAT	TATATTATGA	GTTGGCATCA	GCAAAATCAT	TGGTGTAGTT	GACTGTAGTT	660
30	GCAGATTTA	TAATAAATG	GTAATTAACG	GTCGATATTA	AAATAACTCT	CATTTCAAGT	720
• .	GGGATTAGA	A CTAGTTATTA	AAAAAATGTA	A TACTTTAAGT	GATTTGATG	CATATAATTT	780
25	AAAGTTTTT	C ATTTCATGCT	TTDTTAAAA 1	ATTATTGTA	A TGTAGACTG	GACTGGAATT	840
35	ATTATAGTG	T AAATTTATGO	C ATTCAGTGTA	A AAATTAAAG	r ATTGAACTT	TCTGTTTTAG	900
	አአ አ ልሞ ል ርጥጥ	т атастттаа:	r ataggattt	r gtcatgcga	A TTTAAATTA	A TCGATATTGA	960

	ACACGGAATA CCAAAATTAA AAAGGATACA CATGGCCTTC ATATGAACCG TGAACCTTTG	1020
	ATAACGTGGA AGTTCAAAGA AGGTAAAGTT TAAGAATAAA CTGACAAATT AATTTCTTTT	1080
5	ATTTGGCCCA CTACTAAATT TGCTTTACTT TCTAACATGT CAAGTTGTGC CCTCTTAGTT	1140
·	GAATGATATT CATTTTCAT CCCATAAGTT CAATTTGATT GTCATACCAC CCATGATGTT	1200
.0	CTGAAAAATG CTTGGCCATT CACAAAGTTT ATCTTAGTTC CTATGAACTT TATAAGAAGC	1260
	TTTAATTTGA CATGTTATTT ATATTAGATG ATATAATCCA TGACCCAATA GACAAGTGTA	. 1320
	TTAATATTGT AACTTTGTAA TTGAGTGTGT CTACATCTTA TTCAATCATT TAAGGTCATT	1380
15	AAAATAAATT ATTTTTTGAC ATTCTAAAAC TTTAAGCAGA ATAAATAGTT TATCAATTAT	1440
	TAAAAACAAA AAACGACTTA TTTATAAATC AACAAACAAT TTTAGATTGC TCCAACATAT	1500
20	TTTTCCAAAT TAAATGCAGA AAATGCATAA TTTTATACTT GATCTTTATA GCTTATTTTT	1560
	TTTAGCCTAA CCAACGAATA TTTGTAAACT CACAACTTGA TTAAAAGGGA TTTACAACAA	1620
	GATATATATA AGTAGTGACA AATCTTGATT TTAAATATTT TAATTTGGAG GTCAAAATTT	1680
25	TACCATAATC ATTTGTATTT ATAATTAAAT TTTAAATATC TTATTTATAC ATATCTAGTA	1740
	AACTTTTAAA TATACGTATA TACAAAATAT AAAATTATTG GCGTTCATAT TAGGTCAATA	1800
30	AATCCTTAAC TATATCTGCC TTACCACTAG GAGAAAGTAA AAAACTCTTT ACCAAAAATA	1860
•	CATGTATTAT GTATACAAAA AGTCGATTAG ATTACCTAAA TAGAAATTGT ATAACGAGTA	1920
	AGTAAGTAGA AATATAAAAA AACTACAATA CTAAAAAAAA TATGTTTTAC TTCAATTTCG	1980
35	AAACTAATGG GGTCTGAGTG AAATATTCAG AAAGGGGAGG ACTAACAAAA GGGTCATAAT	2040
	GTTTTTTTAT AAAAAGCCAC TAAAATGAGG AAATCAAGAA TCAGAACATA CAAGAAGGCA	2100

	GCAGCTGAAG CAAAGTACCA TAATTTAATC AATGGAAATT AATTTCAAAG TTTTATCAAA	2160
<u>.</u>	ACCCATTCG	2169
5	(2) INFORMATION FOR SEQ ID NO: 15:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 1165 base pairs	
10	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
15		
	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: YES	-
20		
•		,÷
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:	
25	CTGTCAAAGA AATTCTCGAG GTTACATGGA TATCTTGAGA ACTTAAGAAA TTTTACAGTA	60
	TAATTGAACA AGTATATGCA GCATATCCTA ATTTCTGGAC TGACTGGTAG CCATAAACTG	120
20	AATTTGAATT CATAGAAATT ATTGGAGTAG CGTTTGAGCT TCTCAAGGTC CATACAAAGA	.180
30	ACACATTCTC AACTATCCGT CTCATAGGAT ACAACATTTT CAATTGCAGT TCAACACCAA	240
•	AAAAATGTAA AAAATAGAAA CATCATGACC AGGTAATCAA AACATACTCG TTCGATACGG	300
35	AATCTATTAT TGGTACATTT AAAAGGCTAG AAAAAACAAA CTTCAGTAGC TATCTCAGCA	360
	TO THE TRANSPORTE AGENTANCE ATTACANT THATATACT TTCACANAGA	4,20

	AACAATTTTT	ATCATATCCC	TGGACATATA	ATGAACCCTT	TATGTGTTCA	GAACTTTGCC	480
	CTTGACCATG	TATTTGTGTT	GTAAAAAATC	CACTTATTAT	GTATACATAA	TTGATTTACA	540
5	ACAACAAACA	CAATGTAATC	CCACAAGTGG	AGTGTGGTGA	GGACTTTACC	CCTACCTTAC	600
	GAGATAGAGA	GATTGTTTCT	AATAGACCCT	CGGCTAAAGT	AAAAGCATTT	CAAAGCAACG	660
	ССААТАТААА	GAAGGCATGA	TAAAACACTA	AAGGAAGCAT	GCTAGAGCAT	TCTTACCGAG	720
.0	GAACAATAAC	TACGACAAGA	ТАТАТААТАС	AATAATCGAA	GTACAAGAAA	CAGAAAATAG	780
	AATAACAAAG	ATCAAATAAC	AAAACAAGAA	ACTACCCAAA	TAATTCCACG	ACTACTAGTA	840
15	TGAAAGGATA	AGCCAGACAA	CACTCAAATA	CCTAACTAAC	CTTCTACCCC	TCATCCGTGT	900
	CCTCCATAAC	CTCCTAGAAC	ACTCTTTCTA	AATATTGTCT	TCCCCCACCC	CCCCTCCATC	960
	TCTCAATTT	TGAATTTTAT	' ACACTCAACO	: ACCTTGCAA	A TTTGTCACAI	GATACTTACA	1020
20	TATGGCTCT	A CAAGTGTCAT	TTTTCTTCC	A TATTTGATA	CAAAAATAT 1	AAAATAAAA 1	1080
	ACTAAGGAGA	A TGATCCAGAT	TATATTGGAA	A ATGAAATGC	A AAGGCTAAA	A ATAATTGAAA	1140
25	TTAACATGA	A ATTAGTAAA	ATTAC				1165

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

30

(A) LENGTH: 317 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

	GCAAGCAATG	CACCACAGTT	AGTTTATATC	AAAAAGAAGA	AAGGTATTAA	CGGAGCTAAA	. 60
0	AACTGTTATA	TACCACATGA	AAGAAGTTGA	TAATGTGAAA	ACACCATGCT	CATAAAGATT	120
	GTAATTCAAA	TAACAAATGC	CCACAGGAGT	AAAGAGCTGT	CTTTCCCAAG	TTAAGGTATT	180
	ATAAATTGGC	GGAACGAAGT	AACACATGTT	TGACATCTCC	ACACGGTGCA	CAGATCAAAT	240
15	ATGCCATGAG	CACCAGTCCA	GAAGTTTTCC	AACTATTTAT	ATACTATCCA	TGCAACCATA	300
	TAAATTATCA	AACATAC					317

20 (2) INFORMATION FOR SEQ ID NO: 17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 504 base pairs
 - (B) TYPE: nucleic acid
- 25 (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
- 30 (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: YES

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

	CTGCAAAAAA	AGAGAGCAGT	TTACACAAGA	AAAAACTGCT	AAATCTCAAC	AAAAGTATCA	60
,	TGAATTTAAT	ATTAAGGAAG	CTATTTCGAA	CAGAAAGAGT	AACTCATGAT	AATAGAAGGA	120
5	AATTGTGAAG	CAACAGAAGG	AAGACTTTCT	TTATTTCTAC	AAAATTGCTT	TAAGACTATA	180
	TTTGATGCTT	GTATAGTACA	TGTTGAATCC	CCTCAGCTTC	TTTATGTCTA	TACTTTTTT	240
	ATATTTTGAA	TCTCCTTAGT	GAAAATCTTT	GCTTTGCCAC	TGACACTCCG	GGGGTGTGTC	300
.0	ACTTCTCCAA	AAACCTTGTC	TACTTTTTTG	AAGACCCAAT	CAAACAGCTT	TTTAAAAGAT	360
	CAAAAAAATG	GCCAGGTGCC	ACCTAAATGG	AGCCACTACT	TACTCCCCGG	TATGCAAAAT	420
15	TCTCTAGCAA	AGTCAAAGTA	GGTATAAACA	ATTCATCTTC	CAAAATAAGG	TCAAACTGCC	480
	TAAAGCACAA	CTTTTGGCTG	TTAC				504

(2) INFORMATION FOR SEQ ID NO: 18:

20

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 146 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
- 25 (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO

30

(iv) ANTI-SENSE: YES

35

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

٠.	AGCCAATGAT GCTGCAACAT CATGCTTTAA TAGGAAAATC TGTTATGATG ATGGAAACTA	120
	CTATTTTGTA GTAGACGAGG ACCTAC	146
5		
	(2) INFORMATION FOR SEQ ID NO: 19:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 218 base pairs	
10	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
. *	(D) TOPOLOGY: linear	
,	(ii) MOLECULE TYPE: DNA (genomic)	
15	\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	
15	(iii) HYPOTHETICAL: NO	
	(111)	
	(iv) ANTI-SENSE: YES	•
20		
•		*
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:	
25	CTGTTTAATT GCTGAAGTAG TAAGTTCTCA AGCACTTATA GAATTGACTC ATTTTGTTAA	60
	GGGAAAGAGT ATGGGATCAA GTCCAAATTA GTAAAGACAC AATTATTTTA ACTTTTGCAT	120
	TTCAAAATGT CTTACATAAC AAGACTAGTA AGAACATGAA TCGAAATGCC TGTGATGATG	180
30		210
	GTGTTCAAAA TTCAGCTTCA AGGTATGAAT AACAAAAC	218
	(2) INFORMATION FOR SEQ ID NO: 20:	
35	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 198 base pairs	
•	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	

	50	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
5	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: YES	
10		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:	
	CTGTATCCAG CAGACATAAT AGGAGTGAAC ATAAAAATGT CACTGGATAA ATAACTTATC	60
15	ATGATATTCA GCGGCTACCA ATATTCTGAA GGCCCATGGC GAAAATAAGT ACTTTTATAC	120
	TTTCAGGACG TATATATTTG GATTCTATCT AACAATTGTT CTGAGAATTA TTTAGTTGTA	180
20	GAAATAAATT TAAAATAC	198
	(2) INFORMATION FOR SEQ ID NO: 21:	
	(i) SEQUENCE CHARACTERISTICS:	•
25	(A) LENGTH: 208 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
30	(ii) MOLECULE TYPE: DNA (genomic)	
	(iii) HYPOTHETICAL: NO	

(iv) ANTI-SENSE: YES

293

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(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:	21:
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	(X1) SEQUENCE DESCRIPTION: SEQ ID NO: 21:	
٠.	CTGTGGTTAG AAGCTAAAAG TGAATAGATG AGAAAAATTA CCTCCAAATA AGAGGGATAT	60
5	TGAAAAAGAA ACACAATGCA TGAAAAGAAT AAACAAATGA TAAACGAGAA AATTGAATAA	120
	TCCATCAGAA CCCTGGTTAC CTCACAAAGA GTGAGATTTT CCGTGGCTAA CCTATATGAA	180
0	CCTTAAAATG CAATAGAAAC AGACAAAC	208
	(2) INFORMATION FOR SEQ ID NO: 22:	
	(i) SEQUENCE CHARACTERISTICS:	
5	(A) LENGTH: 293 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
.0	(iii) HYPOTHETICAL: NO	· ·
	(iv) ANTI-SENSE: YES	
25		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 22:	
30	CTGTACAAGT TCATCAAACA TTTCACAATT ACTCCAAAAC AGACACACTT GCAAACTCTA	60
•	TACAGTAATC TTCTATACTA CAAAAAAGTA AACAATGTTT TTTTTAAGAT GACATTTGTT	120
	CTCAGCAACA TAATAGAAAT CCCTAGACAA TGGAAACATT CATCATGTTG TTTTCCTCTA	180

TGTTTCAACC CCTTTGATGT TCAACAGTTC AGGTCATTTT GAGGAATGAA TCTTGTTCAA

GTAAGCCAAA CTAATTGTAA TTATCACAAA ATATCTAAAG ATGTAAGACA TAC

(2)	INFORMATION	FOR	SEQ	ID	NO:	23:
121	TIME OF GRANT TOTAL		~-×			

(i) SEQUENCE CHARACTERISTICS:

5 (A) LENGTH: 376 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: YES

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 23:

20

20	CTGCATTTCA	TCATGAGGGG	GAGGAAAGAC	GGAGAAATAT	AGATATCAGA	TTTAGACCAT	60
	TTCAATTAGT	ATCACTTCAT	TGTAAAGAAA	AGGTAAGTAT	CCAACAAATA	TAGCAGGCTG	120
25	TGGATTGGTA	GCCTGAAACT	ATAGCTTCAA	AGAATCAACT	TAAGCTGCTC	ATCAAGGCCT	180
	TAGTGGTAGA	AATGAGGCGG	TAATAAGTGT	AAATGAATCT	AATACTTGGA	TCTCGAAACA	240
	AAAATCAGAA	ATTCGGTTGG	AAAATAAGTA	GAACAAGATG	AAATGAGCTA	TCATCCCCAG	300
30	AACCAAGTAG	ACTTCCAAGT	AAGCAATCTA	AAAATTACTA	GATTATTTAA	CAAGCTGCGA	360
	TTCAAAATAC	TTGAAC					376

- 35 (2) INFORMATION FOR SEQ ID NO: 24:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 172 base pairs

	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
,	(D) TOPOLOGY: linear	
5	(ii) MOLECULE TYPE: DNA (genomic)	
	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: YES	
10		
		•
•		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 24:	
15		
	CTGCAAAGTG AAGTAACTAA TCAGTACAGC TATTACCGAA TTTGACCAGC TATTGGATTA	60
		· .
	AATAATATGA AATCCATCAT CAAGAAATGG AAGGTAAAAA GGTTTCTACT TGTCCTTGGA	120
20	TAGAATTAAA GCACTTCATA AACCCAACAC TTTCAACTTT AGATGATTTT AC	172
	(2) INFORMATION FOR SEQ ID NO: 25:	
		٠
•	(i) SEQUENCE CHARACTERISTICS:	•
25	(A) LENGTH: 145 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
		٠
30	(ii) MOLECULE TYPE: DNA (genomic)	
		•
•	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: YES	,
35		

54

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:	25:

	CTGTTTTCGT CATGCGAGGA TCAGAAAAAA GAGTTAAATT AGACAATGTG AAAATGATTT	60
5	GTTTCAGTTA CTTCTCCATA AAACTTGTTC AGTACATTAA AAACAAGCAG AGCAATAATT	120
	TCATGGATAA GTAAAACATA TATAC	145
10	(2) INFORMATION FOR SEQ ID NO: 26:	•
10	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 242 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
15	(D) TOPOLOGY: linear	
13	(b) lorobodi. linear	
	(ii) MOLECULE TYPE: DNA (genomic)	
	(II) Monneoud III . Dis. (genemic)	
	(iii) HYPOTHETICAL: NO	
20		
	(iv) ANTI-SENSE: YES	
25		·
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26:	
.•	CTGTCATGAG AACAGATTGT ATGTCAGCAT GAAGACAAAG ATCATCAATA AACAGTTTTC	60
30	TCCTTTTTGA ATTAGCTAAA CAACGCAGGG GGAGGGCAGG AGGCTCAAAC ACTTCCGAAC	120
•	TCAGACAGTC GGATATCTTA TACAACTAAA GATGGATGAG ACAATTACAG TTCTTTTTGG	180
25	TGAGAGAACT GTACCCTACA TCTGTTATCT TATTATCAAA AGTTATTCAA GCAAATCCTT	240

(2) INFORMATION FOR SEQ ID NO: 27:

AC

PCT/IB98/00295

(i) SEQUENCE CHARACTERISTIC

(A) LENGTH: 797 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

10 (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 27:

	CTTCACAAAC	AAGGAGAAGA	AGAAGCAAAA	AGAAAGATGA	ATATAGTTAG	CTTAGTTCAA	60
20	TATAAAAAT	TTCTCTCCAA	GCTATTTTC	TGCTAGCAAA	ATTCATTAGT	TATTTAACTT	120
	TTCTATACAT	AAAGCTGCAC	AAAGAAATAG	TAGTACATTT	TTTTGACTTG	CACAAAATAA	180
25	CTGTGTTGTC	CATTTTCTGA	CATGTGTTCA	TCTACATGCA	CTGTTTCAAC	AACAACAACT	240
	ACTTCAGTCC	CAAACAAGTT	GGGTCGCTTT	AGCTACACAT	GTTGCTTTCA	CTTCTGTTAC	300
30 .	TTCTTTTTGG	ACTTTTTTC	TTGAGCCAAG	GGTCTATTGA	AAAAATCCTC	TCTACCTCTG	360
30 .	AGATAGGAGT	AAGTTTTGCA	TACACTCTAC	сстссссств	AAACCACTTT	GTGGGACTAC	420
•	ACGAGGTATG	TTGTTGTTGA	TGTTAGCGCA	GACACCAAAG	GTGGACATTA	TATGACTATT	480
35	CCTAGCTTTA	CTTCAGGGCG	GTTTTAAGTT	CCCATCAACT	TCATTTTTGA	TCATTTACCT	540
	AAGTTTATGC	AGGTGCAAGC	TACATGCACT	GGTTTAGGGA	AAAAGAGGAT	AGAGAAGAAT	600

PCT	/TRQR	/00295

	·	
	TTTTTTGGCA TCCTTTTGTT TTGTAACAGT AAGATGCCAA AAGTAGACCT TATTACGGCT	660
	ATTCCTACCT TTCAAATTAG TAGTTCAGAG GACTTAACTG GCGATTGTGG CGGTAATCAA	720
5	TAGTTAACTT CTATCGCATT CAAATAACTA TGAACAAAAC CACAATAAAA AGGGAGGTCA	780
	CACGGCAAGA ACTGTAC	797
D	(2) INFORMATION FOR SEQ ID NO: 28:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 2169 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	٠.
5	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: DNA (genomic)	•
	(iii) HYPOTHETICAL: NO	
0	(iv) ANTI-SENSE: YES	
25		
-	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 28:	
•	CGAATGGGTT TTGATAAAAC TTTGAAATTA ATTTCCATTG ATTAAATTAT GGTACTTTGC	60
30	TTCAGCTGCT GCCTTCTTGT ATGTTCTGAT TCTTGATTTC CTCATTTTAG TGGCTTTTTA	120
	TAAAAAAACA TTATGACCCT TTTGTTAGTC CTCCCCTTTC TGAATATTTC ACTCAGACCC	180
•	CATTAGTTTC GAAATTGAAG TAAAACATAT TTTTTTTAGT ATTGTAGTTT TTTTATATTT	240
35	CTACTTACTT ACTCGTTATA CAATTTCTAT TTAGGTAATC TAATCGACTT TTTGTATACA	300
	TACTOR ASSESSMENT TACTOR TACTOR TAGTGGTAAG GCAGATATAG	360

	TTAAGGATTT	ATTGACCTAA	TATGAACGCC	ATTTTAATAA	TATTTTGTAT	ATACGTATAT	420
	TTAAAAGTTT	ACTAGATATG	TATAAATAAG	АДАДТТАТА	TTTAATTATA	AATACAAATG	480
5	ATTATGGTAA	AATTTTGACC	TCCAAATTAA	AAATTTATAA	ATCAAGATTT	GTCACTACTT	540
	АТАТАТАТСТ	TGTTGTAAAT	CCCTTTTAAT	CAAGTTGTGA	GTTTACAAAT	ATTCGTTGGT	600
0	TAGGCTAAAA	AAAATAAGCT	ATAAAGATCA	AGTATAAAAT	TATGCATTTT	CTGCATTTAA	660
. •	TTTGGAAAAA	TATGTTGGAG	CAATCTAAAA	TTGTTTGTTG	ATTTATAAAT	AAGTCGTTTT	720
	TTGTTTTTAA	TAATTGATAA	ACTATTTATT	CTGCTTAAAG	TTTTAGAATG	TCAAAAATA	780
15	ATTTATTTA	ATGACCTTAA	ATGATTGAAT	AAGATGTAGA	CACACTCAAT	TACAAAGTTA	840
	CAATATTAAT	ACACTTGTCT	ATTGGGTCAT	GGATTATATC	: ATCTAATATA	AATAACATGT	900
20	CAAATTAAAG	CTTCTTATAA	AGTTCATAGG	AACTAAGATA	AACTTTGTGA	ATGGCCAAGC	960
	ATTTTTCAGA	ACATCATGGG	TGGTATGACA	ATCAAATTGA	ACTTATGGGA	A TGAAAAATGA	1020
0.5	ATATCATTCA	\ ACTAAGAGGG	CACAACTTGA	. CATGTTAGAA	A AGTAAAGCAA	A ATTTAGTAGT	1080
25	GGGCCAAATA	A AAAGAAATTA	ATTTGTCAGT	TTATTCTTAL	A ACTTTACCT	r CTTTGAACTT	1140
	CCACGTTATO	AAAGGTTCAC	: GGTTCATATG	AAGGCCATG	r gtatccttt	r TAATTTTGGT	1200
30	ATTCCGTGTT	r CAATATCGAT	TAATTTAAAT	TCGCATGAC	A AAATCCTAT	A TTAAAGTATA	1260
٠.	AAGTATTTT	TAAAACAGAG	AAGTTCAATA	CTTTAATTT	T ACACTGAAT	G CATAAATTTA	1320
	CACTATAAT	A ATTCCAGTC	G CAGTCTACAT	TACATAAT	T AACAATTTT	A GCATGAAATG	.1380
35	AAAAACTTT	A AATTATATG	CATCAAATC	A CTTAAAGTA	T ACATTTTT	T AATAACTAGT	1440
	መረመን አጥርርር	አ ሮሞሞሮኔ አልሞሮ	ል ር ልርፕፕል ፕ ፕ	r aatatcgac	C GTTAATTAC	C ATTTTATTAT	1500

	TAAATCTGCA	ACTACAGTCA	ACTACACCAA	TGATTTTGCT	GATGCCAACT	CATAATATAA	1560
	TATCCACCGT	TCATGTGATT	AATTCAATAT	TTCATATACG	TACGTAAÇAA	AAATTACTAA	1620
5	ATTAACGTTG	GATATACCAT	ACCCTAAGCT	CTGCCAAATG	TCAATGTTCT	ATCATTAGCT	1680
	ATTTTTATGC	АТСТАТААТА	GATGTTAAAT	TCATATTCTA	AGATTGAACT	TAATCATAAA	1740
10	CTCAAAATTT	GTGGTACCTG	TCAATGCCTC	CAAAAGTTGA	TTGAACATAA	ACGTTAAGAT	1800
	CTGTGTACTT	GTCTTTTCCT	TGTAATAATG	TATGTATGAT	AATAATAATA	AGAGAACAAA	1860
	ATATGGCAAA	ATAAACACTT	TTTTAACATG	TAACTCAAAA	CAAGTAATAG	GCAAAAGTAC	1920
15	AGATGACAAC	ACAACACTGT	AAACATCATT	GAGGAAAACA	AAAACCATAC	AACATTTTGA	1980
	CTGTAAATGA	AGAGTTTGAA	AACAAAAACT	ATGTTCAAAC	CGACGCCAAG	CTAACGAAAA	2040
20	TAGCCATAGA	. GTTCTAAGAA	GCAGATGCAA	CAGTTCCACG	GGTTAGTATC	GTCTGTAGTA	2100
	GGACCGGTCA	TGAGAACTCG	AAAGAATCTG	AAAGGAAGTA	ATGCATTTGA	ACCAGTAATT	2160
	GGCCATGAT						2169

30

(2) INFORMATION FOR SEQ ID NO: 29:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 11469 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

35

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 29:

	ATCATGGCCA	ATTACTGGTT	CAAATGCATT	ACTTCCTTTC	AGATTCTTTC	GAGTTCTCAT	60
	GACCGGTCCT	ACTACAGACG	ATACTAACCC	GTGGAACTGT	TGCATCTGCT	TCTTAGAACT	120
.0	CTATGGCTAT	TTTCGTTAGC	TTGGCGTCGG	TTTGAACATA	GTTTTTGTTT	TCAAACTCTT	180
	CATTTACAGT	CAAAATGTTG	TATGGTTTTT	GTTTTCCTCA	ATGATGTTTA	CAGTGTTGTG	240
15	TTGTCATCTG	TACTTTTGCC	TATTACTTGT	TTTGAGTTAC	ATGTTAAAAA	AGTGTTTATT	300
•	TTGCCATATT	TTGTTCTCTT	ATTATTATTA	TCATACATAC	ATTATTACAA	GGAAAAGACA	360
	AGTACACAGA	TCTTAACGTT	TATGTTCAAT	CAACTTTTGG	AGGCATTGAC	AGGTACCACA	420
20	AATTTTGAGT	TTATGATTAA	GTTCAATCTT	' AGAATATGAA	TTTAACATCT	ATTATAGATG	480
	САТАААААТА	GCTAATGATA	GAACATTGAC	: ATTTGGCAGA	GCTTAGGGTA	TGGTATATCC	540
25	AACGTTAATT	TAGTAATTT	TGTTACGTAC	GTATATGAAA	. TATTGAATTA	ATCACATGAA	600
	CGGTGGATAI	TATATTATGA	A GTTGGCATC	A GCAAAATCAT	TGGTGTAGT	GACTGTAGTT	660
	GCAGATTTAA	A TAATAAATO	GTAATTAACO	GTCGATATTA	AAATAACTCI	CATTTCAAGT	. 720
30	GGGATTAGA	A CTAGTTATT	A AAAAATGT	A TACTTTAAGI	GATTTGATGO	CATATAATTT	780
	AAAGTTTTT	C ATTTCATGC	r aaaattgtt	A ATTATTGTA	A TGTAGACTG	GACTGGAATT	840
35	ATTATAGTG	r aaatttatg	C ATTCAGTGT	A AAATTAAAG	r attgaactto	G TCTGTTTTAG	900
	AAAATACTT	I ATACTTTAA	T ATAGGATTT	T GTCATGCGA	A TTTAAATTA	A TCGATATTGA	9,60

	ACACGGAATA	CCAAAATTAA	AAAGGATACA	CATGGCCTTC	ATATGAACCG	TGAACCTTTG	1020
	ATAACGTGGA	AGTTCAAAGA	AGGTAAAGTT	TAAGAATAAA	CTGACAAATT	AATTTCTTTT	1080
5	ATTTGGCCCA	CTACTAAATT	TGCTTTACTT	TCTAACATGT	CAAGTTGTGC	CCTCTTAGTT	1140
	GAATGATATT	CATTTTTCAT	CCCATAAGTT	CAATTTGATT	GTCATACCAC	CCATGATGTT	1200
10	CTGAAAAATG	CTTGGCCATT	CACAAAGTTT	ATCTTAGTTC	CTATGAACTT	TATAAGAAGC	1260
ıu	TTTAÄTTTGA	CATGTTATTT	ATATTAGATG	ATATAATCCA	TGACCCAATA	GACAAGTGTA	1320
	TTAATATTGT	AACTTTGTAA	TTGAGTGTGT	CTACATCTTA	TTCAATCATT	TAAGGTCATT	1380
15	TTAAATAAAA	ATTTTTTGAC	ATTCTAAAAC	TTTAAGCAGA	ATAAATAGTT	TATCAATTAT	1440
	ТАААААСААА	. AAACGACTTA	TTTATAAATC	AACAAACAAT	TTTAGATTGC	TCCAACATAT	1500
20	TTTTCCAAAT	' TAAATGCAGA	AAATGCATAA	TTTTATACTI	GATCTTTATA	GCTTATTTTT	1560
	TTTAGCCTAA	CCAACGAATA	TTTGTAAACI	CACAACTTGA	A TTAAAAGGGA	TTTACAACAA	1620
•	GATATATATA	AGTAGTGACA	AATCTTGATT	TTAAATATT	TAATTTGGAG	GTCAAAATTT	1680
25	TACCATAATO	ATTTGTATT	CAAATTAATA	TATAAATTT	TTATTTATAC	ATATCTAGTA	1740
	AACTTTTAAA	A TATACGTATA	A TACAAAATAT	TTATTAAAA 1	GCGTTCATA	T TAGGTCAATA	1800
30	AATCCTTAAC	TATATCTGC	TTACCACTAC	g gagaaagta	A AAAACTCTT	r accaaaata	1860
	CATGTATTA	r gtatacaaa	A AGTCGATTA	G ATTACCTAA	A TAGAAATTG	T ATAACGAGTA	1920
	AGTAAGTAG	AAAATATAA A	A AACTACAAT	а стааааааа	A TATGTTTA	C TTCAATTTCG	1980
35	AAACTAATG	G GGTCTGAGT	G AAATATTCA	g aaaggggag	G ACTAACAAA	A GGGTCATAAT	2040
	GTTTTTTTA	T AAAAAGCCA	C TAAAATGAG	G AAATCAAGA	A TCAGAACAT	A CAAGAAGGCA	2100

	GCAGCTGAAG	CAAAGTACCA	TAATTTAATC	AATGGAAATT	AATTTCAAAG	TTTTATCAAA	2160
٠.	ACCCATTCGA	GGATCTTTTC	CATCTTTCTC	ACCTAAAGTT	TCTTCAGGGG	TAATTTTAC	2220
5	TAATTTCATG	TTAATTTCAA	TTATTTTAG	CCTTTGCATT	TCATTTTCCA	ATATATCTGG	2280
	ATCATCTCCT	TAGTTTTTTA	TTTTATTTT	TATAATATCA	AATATGGAAG	AAAAATGACA	2340
	CTTGTAGAGC	CATATGTAAG	TATCATGTGA	CAAATTTGCA	AGGTGGTTGA	GTGTATAAAA	2400
W	TTCAAAAATT	GAGAGATGGA	GGGGGGGTGG	GGGAAGACAA	TATTTAGAAA	GAGTGTTCTA	2460
	GGAGGTTATG	GAGGACACGG	ATGAGGGGTA	GAAGGTTAGT	TAGGTATTTG	AGTGTTGTCT	2520
15	GGCTTATCCT	TTCATACTAG	TAGTCGTGGA	ATTATTTGGG	TAGTTTCTTG	TTTTGTTATT	2580
	TGATCTTTGT	TATTCTATTT	TCTGTTTCTT	GTACTTCGAT	TATTGTATTA	TATATCTTGT	2640
20	CGTAGTTATT	GTTCCTCGGT	AAGAATGCTC	TAGCATGCTT	CCTTTAGTGT	TTTATCATGC	2700
	CTTCTTTATA	. TTCGCGTTGC	TTTGAAATGO	TTTTACTTTA	GCCGAGGGTC	TATTAGAAAC	2760
	AATCTCTCTA	TCTCGTAAGG	TAGGGGTAA	A GTCCTCACCA	CACTCCACTI	GTGGGATTAC	2820
25	ATTGTGTTTG	TTGTTGTAAA	TCAATTATGI	TATACATAAT	AGTGGATTT	TTACAACACA.	2880
	AATACATGGI	CAAGGGCAAA	GTTCTGAAC	A CATAAAGGGT	TCATTATATO	; TCCAGGGATA	2940
30	TGATAAAAAT	TGTTTCTTTG	G TGAAAGTTA	r ataagattto	TTATGGCTT	TGCTGGAAAC	3000
	ATAATAAGTT	T ATAATGCTGA	GATAGCTAC	r gaägtttgt:	TTTTCTAGC	TTTTAAATGT	3060
٠.	ACCAATAATA	A GATTCCGTAT	CGAACGAGT	A TGTTTTGAT	r acctggtca	GATGTTTCTA	3120
35	TTTTTTACA	r ttttttggto	TTGAACTGC	A ATTGAAAAT(G TTGTATCCT	A TGAGACGGAT	3180
	AGTTGAGAA'	r grgttcttt	TATGGACCT	T GAGAAGCTC	A AACGCTACT	C.CAATAATTTC	3240

	TATGAATTCA	AATTCAGTTT	ATGGCTACCA	GTCAGTCCAG	AAATTAGGAT	ATGCTGCATA	3300
	TACTTGTTCA	ATTATACTGT	AAAATTTCTT	AAGTTCTCAA	GATATCCATG	TAACCTCGAG	3360
5	AATTTCTTTG	ACAGGCTTCT	AGAAATAAGA	TATGTTTTCC	TTCTCAACAT	AGTACTGGAC	3420
	TGAAGTTTGG	ATCTCAGGAA	CGGTCTTGGG	ATATTTCTTC	CACCCCAAAA	TCAAGAGTTA	3480
٠ ا0	GAAAAGATGA	AAGGGTATGT	TTGATAATTT	ATATGGTTGC	ATGGATAGTA	TATAAATAGT	3540
O	TGGAAAACTT	CTGGACTGGT	GCTCATGGCA	TATTTGATCT	GTGCACCGTG	TGGAGATGTC	3600
	AAACATGTGT	TACTTCGTTC	CGCCAATTTA	TAATACCTTA	ACTTGGGAAA	GACAGCTCTT	3660
15	TACTCCTGTG	GGCATTTGTT	ATTTGAATTA	CAATCTTTAT	GAGCATGGTG	TTTTCACATT	3720
	ATCAACTTCT	TTCATGTGGT	ATATAACAGT	TTTTAGCTCC	GTTAATACCT	TTCTTCTTTT	3780
20	TGATATAAAC	TAACTGTGGT	GCATTGCTTG	CATGAAGCAC	AGTTCAGCTA	TTTCCGCTGT	3840
	TTTGACCGAT	GACGACAATI	CGACAATGGC	CACCCTAGAG	GAAGATGTCA	AGACTGAAAA	3900
	TATTGGCCTC	CTAAATTTGG	ATCCAÁCTTI	GGAACCTTA1	CTAGATCACT	TCAGACACAG	3960
25	AATGAAGAGA	A TATGTGGATO	AGAAAATGCT	CATTGAAAA	A TATGAGGGAC	CCCTTGAGGA	4020
	ATTTGCTCA	A GGTAACAGCO	AAAAGTTGT	G CTTTAGGCA	TTTGACCTT	A TTTTGGAAGA	4080
30	TGAATTGTT	r ATACCTACT	r TGACTTTGC	r agagaattt	r GCATACCGG(G GAGTAAGTAG	4140
30	TGGCTCCAT	TAGGTGGCAG	CTGGCCATT	TTTTGATCT	r ttaaaaagc	r gtttgattgg	4200
	GTCTTCAAA	A AAGTAGACA	A GGTTTTTGG	A GAAGTGACA	C ACCCCCGGA	G TGTCAGTGGC	4260
35	AAAGCAAAG	A TTTTCACTA	A GGAGATTCA	A AATATAAA	A AAGTATAGA	C ATAAAGAAGC	4320
	TGAGGGGAT	T CAACATGTA	C TATACAAGC	A TCAAATATA	G TCTTAAAGC	A ATTTTGTAGA	4380

	AATAAAGAAA	GTCTTCCTTC	TGTTGCTTCA	CAATTTCCTT	CTATTATCAT	GAGTTACTCT	4440
	TTCTGTTCGA	AATAGCTTCC	TTAATATTAA	ATTCATGATA	CTTTTGTTGA	GATTTAGCAG	4500
5	TTTTTTCTTG	TGTAAACTGC	TCTCTTTTTT	TGCAGGTTAT.	TTAAAATTTG	GATTCAACAG	4560
	GGAAGATGGT	TGCATAGTCT	ATCGTGAATG	GGCTCCTGCT	GCTCAGTAGG	TCCTCGTCTA	4620
.0	CTACAAAATA	GTAGTTTCCA	TCATCATAAC	AGATTTTCCT	ATTAAAGCAT	GATGTTGCAG	4680
. ·	CATCATTGGC	TTTCTTACAT	GTTCTAATTG	CTATTAAGGT	TATGCTTCTA	ATTAACTCAT	4740
	CCACAATGCA	GGGAAGCAGA	AGTTATTGGC	GATTTCAATG	GATGGAACGG	TTCTAACCAC	4800
5	ATGATGGAGA	AGGACCAGTT	TGGTGTTTGG	AGTATTAGAA	TTCCTGATGT	TGACAGTAAG	4860
	CCAGTCATTC	CACACAACTC	CAGAGTTAAG	TTTCGTTTCA	AACATGGTAA	TGGAGTGTGG	4920
20	GTAGATCGTA	TCCCTGCTTG	GATAAAGTAT	GCCACTGCAG	ACGCCACAAA	GTTTGCAGCA	4980
	CCATATGATG	GTGTCTACTG	GGACCCACCA	CCTTCAGAAA	GGTTTTGTTA	TTCATACCTT	5040
	GAAGCTGAAT	TTTGAACACC	ATCATCACAG	GCATTTCGAT	TCATGTTCTT	ACTAGTCTTG	5100
25	TTATGTAAGA	CATTTTGAAA	TGCAAAAGTT	AAAATAATTG	TGTCTTTACT	AATTTGGACT	5160
	TGATCCCATA	CTCTTTCCCT	TAACAAAATG	AGTCAATTCT	ATAAGTGCTT	GAGAACTTAC	5220
30	TACTTCAGCA	ATTAAACAGG	TACCACTTCA	AATACCCTCG	CCCTCCCAAA	CCCCGAGCCC	5280
	CACGAATCTA	TGAAGCACAT	GTCGGCATGA	GCAGCTCTGA	GCCACGTGT	AATTCGTATC	5340
	GTGAGTTTGC	AGATGATGTT	TTACCTCGGA	TTAAGGCAAF	TAACTATAA1	ACTGTCCAGT	5400
35	TGATGGCCAT	AATGGAACAT	TCTTACTATO	G GATCATTTGG	ATATCATGT	r ACAAACTTTT	5460
•	TTGCTGTGAG	CAGTAGATAT	GGAAACCCGG	AGGACCTAA	A GTATCTGAT	A GATAAAGCAC	5520

	ATAGCTTGGG	TTTACAGGTT	CTGGTGGATG	TAGTTCACAG	TCATGCAAGC	AATAATGTCA	5580
	CTGATGGCCT	CAATGGCTTT	GATATTGGCC	AAGGTTCTCA	AGAATCCTAC	TTTCATGCTG	5640
5	GAGAGCGAGG	GTACCATAAG	TTGTGGGATA	GCAGGCTGTT	CAACTATGCC	AATTGGGAGG	5700
	TTCTTCGTTT	CCTTCTTTCC	AACTTGAGGT	GGTGGCTAGA	AGAGTATAAC	TTTGACGGAT	5760
0	TTCGATTTGA	TGGAATAACT	TCTATGCTGT	ATGTTCATCA	TGGAATCAAT	ATGGGATTTA	5820
.0	CAGGAAACTA	TAATGAGTAT	TTCAGCGAGG	CTACAGATGT	TGATGCTGTG	GTCTATTTAA	5880
	TGTTGGCCAA	TAATCTGATT	CACAAGATTT	TCCCAGATGC	AACTGTTATT	GCCGAAGATG	5940
15	TTTCTGGTAT	GCCGGGCCTT	GGCCGGCCTG	TTTCTGAGGG	AGGAATTGGT	TTTGTTTACC	6000
	GCCTGGCAAT	GGCAATCCCA	GATAAGTGGA	TAGATTATTT	AAAGAATAAG	AATGATGAAG	6060
20	ATTGGTCCAT	'GAAGGAAGTA	ACATCGAGTT	TGACAAATAG	GAGATATACA	GAGAAGTGTA	6120
	TAGCATATGO	GGAGACCCAT	GATCAGGTAT	TTTAAATTT	TTTCTACAAC	TAAATAATTC	6180
	TCAGAACAAI	TGTTAGATAG	AATCCAAATA	A TATACGTCCI	GAAAGTATA	AAGTACTTAT	6240
25	TTTCGCCAT	GGCCTTCAG	ATATTGGTAC	CCGCTGAAT!	A TCATGATAA	TTATTTATCC	6300
. •	AGTGACATT	r TTATGTTCAC	C TCCTATTATO	TCTGCTGGA	r acagtctat:	r GTTGGTGACA	6360
30	AGACCATTG	ATTTCTCCTA	A ATGGACAAA	G AGATGTATT	C TGGCATGTC	T TGCTTGACAG	6420
30	ATGCTTCTC	TGTTGTTGA	r cgaggaatt	G CGCTTCACA	A GGTTTGTCT	G TTTCTATTGC	6480
	ATTTTAAGG	r tcatatagg	r tagccacgg	A AAATCTCAC	T CTTTGTGAG	G TAACCAGGGT	6540
35	TCTGATGGA	T TATTCAATT	T TCTCGTTTA	T CATTTGTTT	A TTCTTTCA	T GCATTGTGTT	6600
	тсттттса	A TATCCCTCT	T ATTTGGAGG	T AATTTTCT	C ATCTATTCA	C TTTTAGCTTC	666

	TAACCACAGA	TGATCCATTT	TTTCACAATG	GCCTTGGGAG	GAGAGGGGTA	CCTCAATTTC	6720
.*	ATGGGTAACG	AGGTATGTCT	TACATCTTTA	GATATTTTGT	GATAATTACA	ATTAGTTTGG	6780
5	CTTACTTGAA	CAAGATTCAT	TCCTCAAAAT	GACCTGAACT	GTTGAACATC	AAAGGGGTTG	6840
	AAACATAGAG	GAAAACAACA	TGATGAATGT	TTCCATTGTC	TAGGGATTTC	TATTATGTTG	6900
10	CTGAGAACAA	ATGTCATCTT	ААААААААСА	TTGTTTACTT	TTTTGTAGTA	TAGAAGATTA	6960
lU	CTGTATAGAG	TTTGCAAGTG	TGTCTGTTTT	GGAGTAATTG	TGAAATGTTT	GATGAACTTG	7020
	TACAGTTTGG	CCATCCTGAG	TGGATTGACT	TCCCTAGAGA	GGGCAATAAT	TGGAGTTATG	7080
15	ACAAATGTAG	ACGCCAGTGG	AACCTCGCGG	ATAGCGAACA	CTTGAGATAC	AAGGTTCAAG	7140
	TATTTTGAAT	CGCAGCTTGT	TAAATAATCI	AGTAATTTT	R AGATTGCTTA	A CTTGGAAGTC	7200
20	TACTTGGTTC	TGGGGATGAT	AGCTCATTTC	: ATCTTGTTCT	CTTATTTTC	CAACCGAATT	7260
	TCTGATTTT	r GTTTCGAGAT	CCAAGTATTA	GATTCATTI	A CACTTATȚA	C CGCCTCATTT	7320
	CTACCACTA	A GGCCTTGATO	AGCAGCTTA	A GTTGATTCT	r tgaagctati	A GTTTCAGGCT	7380
25	ACCAATCCA	C AGCCTGCTAT	C ATTTGTTGG	A TACTTACCT	r ttctttaca	A TGAAGTGATA	7440
	CTAATTGAA	A TGGTCTAAA	r ctgatatct	A TATTTCTCC	G TCTTTCCTC	C CCCTCATGAT	7500
20	GAAATGCAG'	T TTATGAATG	C ATTTGATAG	A GCTATGAAT	T CGCTCGATG	A AAAGTTCTCA	7560
30	TTCCTCGCA	T CAGGAAAAC	A GATAGTAAG	C AGCATGGAT	G ATGATAATA	A GGTAAAATCA	7620
	TCTAAAGTT	G AAAGTGTTG	G GTTTATGAA	G TGCTTTAAT	T CTATCCAAG	G ACAAGTAGAA	7680
35	ACCTTTTTA	C CTTCCATTT	C TTGATGATG	G ATTTCATAT	T ATTTAATCO	A ATAGCTGGTC	7740
	AAATTCGGT	A ATAGCTGTA	.C TGATTAGTT	A CTTCACTT	G CAGGTTGTT	G TGTTTGAACG	7800

	TGGTGACCTG	GTATTTGTAT	TCAACTTCCA	CCCAAAGAAC	ACATACGAAG	GGTATATATG	7860
	TTTTACTTAT	CCATGAAATT	ATTGCTCTGC	TTGTTTTAA	TGTACTGAAC	AAGTTTTATG	7920
5	GAGAAGTAAC	TGAAACAAAT	CATTTTCACA	TTGTCTAATT	TAACTCTTTT	TTCTGATCCT	7980
	CGCATGACGA	AAACAGGTAT	AAAGTTGGAT	GTGACTTGCC	AGGGAAGTAC	AGAGTTGCAC	8040
o .	TGGACAGTGA	TGCTTGGGAA	TTTGGTGGCC	ATGGAAGAGT	AAGGATTTGC	TTGAATAACT	8100
.0	TTTGATAATA	AGATAACAGA	TGTAGGGTAC	AGTTCTCTCA	CCAAAAAGAA	CTGTAATTGT	8160
,	CTCATCCATC	TTTAGTTGTA	TAAGATATCC	GACTGTCTGA	GTTCGGAAGT	GTTTGAGCCT	8220
15	CCTGCCCTCC	CCCTGCGTTG	TTTAGCTAAT	TCAAAAAGGA	GAAAACTGTT	TATTGATGAT	8280
	CTTTGTCTTC	ATGCTGACAT	ACAATCTGTT	CTCATGACAG	ACTGGTCATG	ATGTTGACCA	8340
20	TTTCACATCA	CCAGAAGGAA	TACCTGGAGT	TCCAGAAACA	A AATTTCAATG	GTCGTCCAAA	. 8400
-	TTCCTTCAA	GTGCTGTCTC	CTGCGCGAAC	ATGTGTGGT	A CAGTTCTTGC	CGTGTGACCT	8460
	CCCTTTTA	TGTGGTTTTG	; TTCATAGTT	TTTGAATGC	S ATAGAAGTTA	ACTATTGATT	8520
25	ACCGCCACAI	A TCGCCAGTTA	AGTCCTCTG	A ACTACTAAT	r tgaaaggtac	G. GAATAGCCGT	8580
	AATAAGGTC	r acttttggc.	A TCTTACTGT	r acaaaacaa	a aggatgccai	A AAAAATTCTT	8640
30	CTCTATCCT	C TTTTTCCCT!	A AACCAGTGC	A TGTAGCTTG	C ACCTGCATA	A ACTTAGGTAA	8700
	ATGATCAAA	A ATGAAGTTGA	A TGGGAACTT	A AAACCGCCC	T GAAGTAAAG	C TAGGAATAGT	8760
	CATATAATG	T CCACCTTTG	G TGTCTGCGC	T AACATCAAC	A ACAACATAC	C TCGTGTAGTC	8820
35	CCACAAAGT	G GTTTCAGGG	G GAGGGTAGA	G TGTATGCAA	A ACTTACTCC	T ATCTCAGAGG	888
	TAGAGAGGA	T TTTTTCAAT.	À GACCCTTGG	C TCAAGAAAA	A AAGTCCAAA	A AGAAGTAACA	8940

	GAAGTGAAAG	CAACATGTGT	AGCTAAAGCG	ACCCAACTTG	TTTGGGACTG	AAGTAGTTGT	9000
	TGTTGTTGAA	ACAGTGCATG	TAGATGAACA	CATGTCAGAA	AATGGACAAC	ACAGTTATTT	9060
5	TGTGCAAGTC	AAAAAAATGT	ACTACTATTT	CTTTGTGCAG	CTTTATGTAT	AGAAAAGTTA	9120
	AATAACTAAT	GAATTTTGCT	AGCAGAAAAA	TAGCTTGGAG	AGAAATTTTT	TATATTGAAC	9180
	TAAGCTAACT	ATATTCATCT	TTCTTTTTGC	TTCTTCTTCT	CCTTGTTTGT	GAAGGCTTAT	9240
10	TACAGAGTTG	ATGAACGCAT	GTCAGAAACT	GAAGATTACC	AGACAGACAT	TTGTAGTGAG	9300
	CTACTACCAA	CAGCCAATAT	CGAGGAGAGT	GACGAGAAAC	TTAAAGATTC	GTTATCTACA	9360
15	AATATCAGTA	ACATTGACGA	ACGCATGTCA	GAAACTGAAG	TTTACCAGAC	AGACATTTCT	9420
	AGTGAGCTAC	: TACCAACAGO	CAATATTGAG	GAGAGTGACG	; AGAAACTTAA	AGATTCGTTA	9480
20	TCTACAAATA	TCAGTAACAT	TGATCAGACI	GTTGTAGTT	CTGTTGAGGA	A GAGAGACAAG	9540
20	GAACTTAAAG	ATTCACCGTC	TGTAAGCATO	ATTAGTGATO	TTGTTCCAGG	TGAATGGGAT	9600
	GATTCAGATC	CAAACGTCTC	GGGTGAGGA	TAGTCAGATO	3 ATTGATCGAC	CCTTCTACCG	9660
25	ATTGGTGAT	GCTATCCTT	G CTCTCTGAG	A AATAGGTGA	GCGAAACAA	A AAATAATTTG	9720
	CATGATAAA	A AGTCTGATT	TATGATCGC	r atcctcgct	C TCTGAGAAA	G AAGCGAAACA	9780
20	AAGGCGACT	C CTGGACTCG	A ATCTATAAG	A TAACAAAGG	C GACTCCTGG	G ACTCGAATCT	9840
30	ATAAGATAA	CAAAGGCAAT	T CCAAGACTT	G AATCTATAA	A AAATTTAGT	T AAGAATGATT	9900
	AACGTCCGA	T CCTAATTCG	A ATCGAGGCA	T CTTACCACT	C CATTGATAA	T TATATAAGTC	9960
35	AATAAGTCA	T ATAAAGTAT	т ааааастаа	A TTGACTTGA	T CGGTCTATC	A AAAATAGATA	10020
	AATTGTGTT	C ATATGTAAC	A TTTTTGTTG	T CACAATTAG	C TTAATTACA	T CTTTCATGTG	10080

	CAATAACAAA	GAAATGATAG	GAATTTAGAG	ATTCCAATTT	TTTTGTTGCC	ACAATTAACT	10140
	TAATTACATC	TTTCATTTGC	AATAACAAAG	AAATGATAGG	AATTTAGAGA	TCCAGTGTCA	10200
5	ATACACAACC	TAGGCCAACA	TCGAAAGCAT	AACTGTAAAC	TCATGCATGA	AGAAATCAGT	10260
	CGTAAAAATG	AATAAATGCG	ACATAAAAAC	AAATTGCATG	TATCATTAAT	GTGACTTAAC	10320
	TACAAGTAAA	AATAAATTTA	ACAAATGTAA	CTTAACTACA	AGTAAAAATA	AATTGCTTCT	10380
10	ATCATTAACA	AACAAACAGA	ATTAAAAAGA	AAAAAACATA	СТАААТСТТА	CCGTCATTCG	10440
	AAAAAAAAA	ATACCAAATT	CATAATGCAA	GGAAAACGAA	ACGCGTCCTG	ATCGGGTATC	10500
15	AACGATGAAA	TGGACCAGTT	GGATCGACTG	CCTGCACAAC	GTTAGGTATG	CCAAAAAAAA	10560
	GAACACGATC	CTTTGCACCC	GTTCGATGAT	' TATCAGTATG	TTCACAAAAA	AAACTTAAGT	10620
20	TCATCCCAGT	GTACAACAGC	CCCAACATCI	GCCCCAAGTA	ACAAAAAACA	ACCAATTTAT	10680
20	CTTATTCTTA	TCTGCCACAA	AATAATCGGT	TTCACACTAT	TCTCTTGTTA	TACAAAATTG	10740
	ACAAGTAGG	AGGAGAGGAG	TCATCCAAAT	r AAACGGTGCA	. CGTTCTTTG!	A GAAAAGTCTT	10800
25	ATTTTTCGT	A AGATCCAATT	TCAACAAAC	TTTCTTCAAC	TCAAAATTC	TGATAGTGTA	10860
	TCTCCTCTCC	G ACGACCTCTT	GCATTGAAC	G ATCTCCGCT	T ATCATGAAA	A GTTGCTTGGA	10920
20	TAACAAGTA	TGCAAGGGGG	GGACAGTAG	C TATTAAGTT	A GTCGGCCCA	A GGAAATGGAG	10980
30	GAGTGATAG	r ctcgaatat7	T ATTCACCTC	T TTAGCATTAG	CCGGTCTGG	C TTTAAGGAGT	11040
	TACGTCTTT	T ACGCTCGCC	A ATTTCTTTT	T TTAGAATGG	T TGGTGTCAA	A ATCGCGAGTT	11100
35	GTGGAAGGT	T CAAGTTACT	C GATTCGTGA	T TTTCAAGTA	T GAGTGGTGA	G AGAGATTCGA	11160
	TATTTTCAC	G AGGTGTATT	C GAGGTCTAG	T AGAACGAAG	G GTGTCACTA	A TGAAAGTTTC	11220

PCT/IB98/00295

	AAGAGTTCAT CATCATCTTC TTCTAGTAGA TTTTCGCTTT CAAATGAGTA TGAAAATTCT	11280
	TCCTCTTTTC TATTGATTTT CTTCATTGTT TTCTTCATTG TTGTGGTTGT TATTGAAAAG	11340
5	AAAGAAAATT TATAACAGAA AAAGATGTCA AAAAAAAGGT AAAATGAAAG AGTATCATAT	11400
	ACTTAAAGAG TTGCGTAGAG ATAAGTCAAA AGAAACAGAA TTATAGTAAT TTCAGCTAAG	11460
0	TTAGAATTC	11469
U	(2) INFORMATION FOR SEQ ID NO: 30:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 26 base pairs	
5	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
-	(D) TOPOLOGY: linear	
	<pre>(ii) MOLECULE TYPE: other nucleic acid (A) DESCRIPTION: /desc = "Synthetic DNA Primer"</pre>	
20	(A) DESCRIPTION: /desc = Synchectic 2	
	(iii) HYPOTHETICAL: NO	
	(iv) ANTI-SENSE: YES	
25		
		•
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 30:	
<i>5</i> 0	GGAATTCCAG TCGCAGTCTA CATTAC	2
	(2) INFORMATION FOR SEQ ID NO: 31:	
		•

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 28 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single

35

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "Synthetic DNA Primer"

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- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: YES

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 31:
- 15 CGGGATCCAG AGGCATTAAG ATTTCTGG

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- (2) INFORMATION FOR SEQ ID NO: 32:
 - (i) SEQUENCE CHARACTERISTICS:

20

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- 25
- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "Synthetic DNA Primer"
- (iii) HYPOTHETICAL: NO
- 30 (iv) ANTI-SENSE: YES
- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 32:

- (2) INFORMATION FOR SEQ ID NO: 33:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: other nucleic acid
- 10 (A) DESCRIPTION: /desc = "Synthetic DNA Primer"
 - (iii) HYPOTHETICAL: NO
 - (iv) ANTI-SENSE: YES

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 33:

20

CGGGATCCGG GGTAATTTTT ACTAATTTCA TG

32

- (2) INFORMATION FOR SEQ ID NO: 34:
- 25 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

30

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "Synthetic DNA Primer"
- (iii) HYPOTHETICAL: NO

35

(iv) ANTI-SENSE: YES

	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 34:	٠.	•
5	CGGGATCCCG TATGTCTCAC TGTGTTTGTG GC		32
	(2) INFORMATION FOR SEQ ID NO: 35:		
	(i) SEQUENCE CHARACTERISTICS:		
10	(A) LENGTH: 32 base pairs		
	(B) TYPE: nucleic acid		
	(C) STRANDEDNESS: single		
	(D) TOPOLOGY: linear		
15	(ii) MOLECULE TYPE: other nucleic acid		
	(A) DESCRIPTION: /desc = "Synthetic DNA Primer"		
	(iii) HYPOTHETICAL: NO		
20	(iv) ANTI-SENSE: YES		
25	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 35:		
	CGGGATCCCC CTACATACAT ATATCAGATT AG		32
30	(2) INFORMATION FOR SEQ ID NO: 36:		•
<i>5</i> U	(i) SEQUENCE CHARACTERISTICS:		
	(A) LENGTH: 28 base pairs		
	(A) DENGIN. 20 Desc Pesse		

(ii) MOLECULE TYPE: other nucleic acid

(B) TYPE: nucleic acid(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

35

(A) DESCRIPTION: /desc = "Synthetic DNA Primer"

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: YES

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 36:

10

CCATCGATAC TTTAAGTGAT TTGATGGC

28

- (2) INFORMATION FOR SEQ ID NO: 37:
- 15 (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

20

- (ii) MOLECULE TYPE: other nucleic acid
 - (A) DESCRIPTION: /desc = "Synthetic DNA Primer"
- (iii) HYPOTHETICAL: NO

25

(iv) ANTI-SENSE: YES

30

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 37:

CGGGATCCTG TTCTGATTCT TGATTTCC

28

- 35 (2) INFORMATION FOR SEQ ID NO: 38:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2122 base pairs

PCT/IB98/00295

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(B)	TYPE: nucleic	acid
(C)	STRANDEDNESS:	single
(D)	TODOLOGY, line	

5 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

10

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 38:

5		*				•	
	GTATGTCTCA	CTGTGTTTGT	GGCTGTGTGT	GTTTTTTCT	CTGTCTTTTT	GTGTTTTGTG	60
	TAATTGGGGC	TCTTTAAAGT	TGGTATTGTG	TATACCCTTT	TGAGTATAGT	CTTTGAGGAA	120
20	GCAAAATGAT	GAATCTTGAT	TGACATTAGT	AAGGGTTGTA	ACTTTTTGAA	GTTTGGTTAG	180
	GTGTAATTGA	GTTTGGCTTG	TGTGTCTGTG	TGTCGAGGTT	ATTTTTTTGG	TTTGTGTTAT	240
	TGGGGATTCT	TAAAAGTTGG	TATTGTGTAT	ACCCTTTTGA	GTATAGTCTT	TGAGGAAGCA	300
25	AAAATGATGA	ATCTTGATTG	GCATTAGTAA	AGGTTGTAGC	TTTTTGAAGT	GTGGTTAGGT	360
	GTAATTGAGT	TTGGCTTGTG	TGTCTGTGTG	TTTTGGAATC	CTGATGTGTG	TCAAGTCCTG	420
30	ATATGGGTCG	AGGTTCTTTC	TTTGGTTTGT	GTAATTGGGG	GTTCTTAAAA	GTTGGTATTA	480
٠.	TGTACCTTTT	' TAAGAATAGT	GTCTGAGAAA	GCAAAATCGA	TGAATTTTGA	TTGACAGCAT	540
	ATTCTTTGAG	: AAAGCAAAAA	ATGGTGAGTT	TTCATGGAGA	AACTTGATTG	ACATTACTAA	. 600
35	AGGTAGCAAC	TTTTTCAACT	CCTGATATGO	GTCAAGGTTC	TTTGTTTGGT	TTGTGTAATT	660
	TGGGGTTCTI	TGAAGTTTTG	: AGAAAGAAAJ	ATTATGATT	TTCATGGAGA	AATTTGATTT	720

٠.	ACATTAATAA	AGGTAGTAGC	TTTTTAAAGT	GTGGTCAGCT	GTAATGAGTT	CAGCTTGGTT	780
.	TAAAGGGGCC	CTACATATGG	TGCTTTCTGG	TGAGATATTT	GTTGCTCCAC	CATACGAGTT	840
5 ·	ATAAGAATCA	TAGTGTTAGG	ATCTTTTTC	TTTTTTTTT	CATTTTTCAC	TTGACTAGCT	900
	ACTAGAGGAG	TGATCTTGAC	GGCGGAAAAT	CTTAGAAAGG	GGAAGGTTGT	TTGCATCAAC	960
lÖ	TGGTGTTATA	TGTGCAAGGA	GACGGGAGAT	GATGTAGATC	ATCTTCTTCT	TCATTGTGGT	1020
ů.	CTTTCCATGA	GGTTATGATG	TGATATGTTT	GAATGGTTTG	GTACTTCTTG	GCTATGCCAA	1080
	GAACTGTGAA	AGAATTGATA	TTCAGTTGGA	AGTGTGGAGT	TGGAAGAGTG	GAAGAATTGA	1140
15	CACTTGGTTC	CATTAGCTTT	AATGTGGGTG	GTGTGGAGAG	AGAGAGAAAT	AGGAGAGCTT	1200
	TTGAGGGGGT	AGAGTTGAGC	TTTCCTCAGT	TGAGAAGTAG	CCTTTGATAT	CTTTTTTTT	1260
20	TTTTTTTGTA	CACCCATAGA	ATTCCCAATT	GTATAGAAGA	TTGGGTGGAG	TTTGTAGAGA	1320
•	ATCATCTTT	GTAGTAGATT	CTTTACCTTT	TGGTATATCO	: ATTGTATACA	A GCCAGGCCTT	1380
0.5	TGACTATGTT	TATGAATGAA	TATACATTAC	TTGAAAAAA	AAGAAGTGAA	A GCCAGTCTGT	1440
25	TGTACCTTTC	TAGACAATGT	TGTTGCAGCA	TCTTGATAA	TCCCTGAAA	A TTGTCTCCCT	1500
	GAAGGAATAC	TTTGGTTGAT	ATTGATTATI	TCTTGGTTT	TTTAATTCG	G TGTTCTTGAA	1560
30	GGCCATTTI	A AATCCTTTG	A CATTGTTAA!	A GGTGTTTAC	A AGTGTTGGT	C TGGGTTTAAA	1620
	AGCACCTCT	I GTATGGTGCT	TTCTGGAGT	ATCTTTCTT	C CTCCAAAAG	A GAAGTTGCAA	1680
	GAATCAGTG'	T GTGTACTTT	TTCTCTTGT	A TGATCAGAT	C TTTTTTCAA	T TTTTCCGTTT	1740
35	TAGTTGATT	T ATCCATATA	G TGAAAGTTG	G TGTCATAGT	T GCTGTTTGT	G GACTTCCTGT	1800
	A A A GTTTT	T TGATATACT	T AAAAAATTG	T CACACAGAA	.G AAAGAGTTT	TTACCATTAC	1860

PCT/IB98/00295

•	GACACITATO						
	ር እ ር እ ርጥ ጥ እ ጥር	тсссссталс	ттсстстсас	TAGTGGTCTT	TAATTGTGGA	GATATAACTA	2100
5	TTGGAGCATC	ACTTCTAATC	ATAAAAGTCT	TTGCTCTCTT	CAACCATGAA	TGATAAATTG	2040
	CGTGTACTTG	AAATAGTTTG	GTAAAATTGT	GATAGGAAAA	AAGATAATTC	TTGATTGCTŢ	1980
	TTAAGCTAGA	TGGGACTGTT	TGATTCTTAG	ACCAAATAAT	GAACCTTTTT	GTTCTCTTAA	1920

CLAIMS

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- 1. A method of affecting enzymatic activity in a plant (or a cell, a tissue or an organ thereof) comprising expressing in the plant (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for an intron of a class A potato starch branching enzyme in a sense orientation, optionally together with a nucleotide sequence which codes, partially or completely, for an intron of a class B starch branching enzyme in a sense or antisense orientation; and wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron.
 - 2. A method according to claim 1 wherein starch branching enzyme activity is affected and/or wherein the levels of amylopectin are affected and/or the composition of starch is changed.
 - 3. A method of affecting enzymatic activity in a starch producing organism (or a cell, a tissue or an organ thereof) comprising expressing in the starch producing organism (or a cell, a tissue or an organ thereof) a nucleotide sequence wherein the nucleotide sequence codes, partially or completely, for an intron of a class A starch branching enzyme in a sense orientation, optionally together with a nucleotide sequence which codes, partially or completely, for an intron of a class B starch branching enzyme in a sense or antisense orientation; wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron; and wherein starch branching enzyme activity is affected and/or the levels of amylopectin are affected and/or the composition of starch is changed.
 - 4. A method according to any one of claims 1 to 3 wherein the nucleotide sequence does not contain a sequence that is sense to an exon sequence.
- 30 5. A method according to any one of the preceding claims wherein the enzymatic activity is reduced or eliminated.

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- 6. A method according to any one of the preceding claims wherein the nucleotide sequence codes for at least substantially all of at least one intron in a sense orientation.
- 5 7. A method according to any one of the preceding claims wherein the nucleotide sequence codes for all of at least one intron in a sense orientation.
 - 8. A method according to any one of the preceding claims wherein the nucleotide sequence comprises the sequence shown as SEQ. ID. No. 38, or a variant, derivative or homologue thereof.
 - 9. A method according to any one of the preceding claims wherein the nucleotide sequence is expressed by a promoter having a sequence shown as SEQ.I.D. No. 14 or a variant, derivative or homologue thereof.
- 10. A promoter having a sequence shown as SEQ.I.D. No. 14, or a variant, derivative or homologue thereof.
 - 11. A promoter according to claim 10 in combination with a gene of interest ("GOI").
 - 12. A construct capable of comprising or expressing the invention according to any one of claims 10 and 11.
- 13. A vector comprising or expressing the invention according to any one of claims 10 to 12.
 - 14. A combination of nucleotide sequences comprising a first nucleotide sequence coding for a recombinant enzyme; and a second nucleotide sequence which corresponds to a class A SBE intron in a sense orientation; wherein the intron is an intron that is associated with a genomic gene encoding an enzyme corresponding to the recombinant

WO 98/37214 PCT/IB98/00295

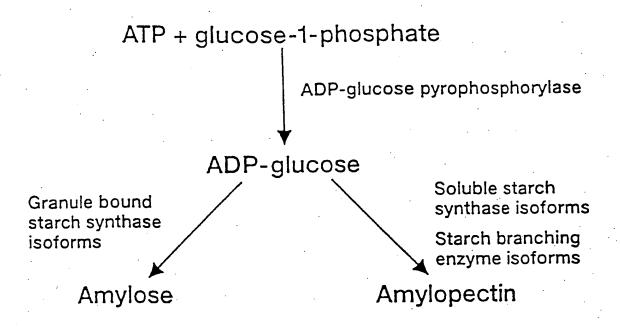
enzyme; and wherein the second nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron.

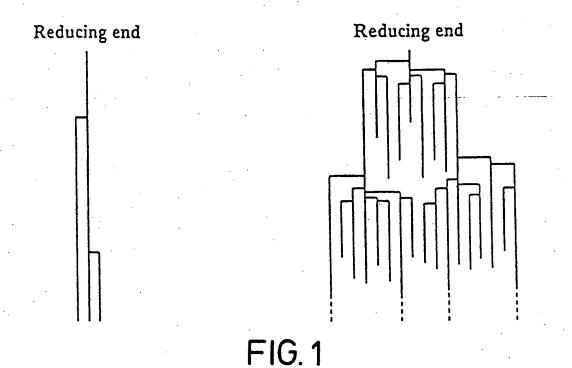
- 15. A cell, tissue or organ comprising or expressing the invention according to any one of claims 10 to 14.
 - 16. A transgenic starch producing organism comprising or expressing the invention according to any one of claims 10 to 15.
- 10 17. A transgenic starch producing organism according to claim 16 wherein the organism is a plant.
 - 18. A starch obtained from the invention according to any one of the preceding claims.

15

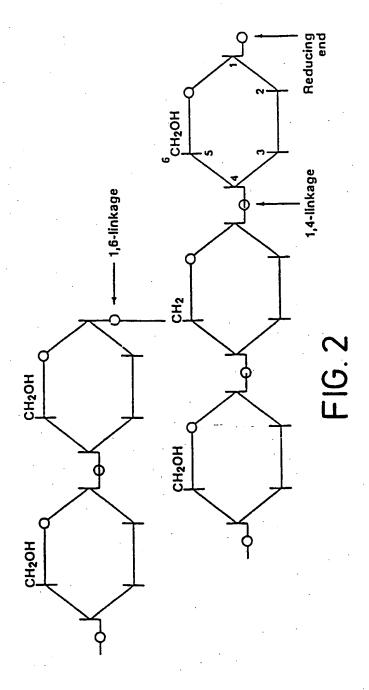
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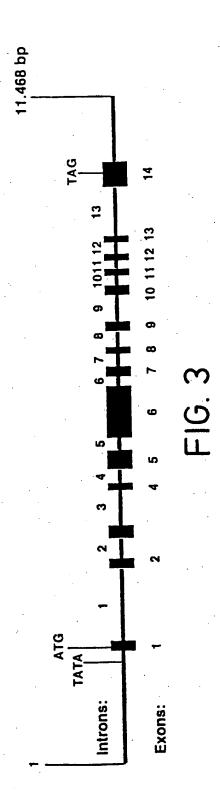
19. A method of expressing a recombinant protein or enzyme in a host organism comprising expressing a nucleotide sequence coding for the recombinant protein or enzyme; and expressing a further nucleotide sequence; wherein the further nucleotide sequence codes, partially or completely, for a class A SBE intron in a sense orientation; wherein the intron is an intron normally associated with the genomic gene encoding a protein or an enzyme corresponding to the recombinant protein or enzyme; and wherein the further nucleotide sequence does not contain a sequence that is sense to an exon sequence normally associated with the intron.



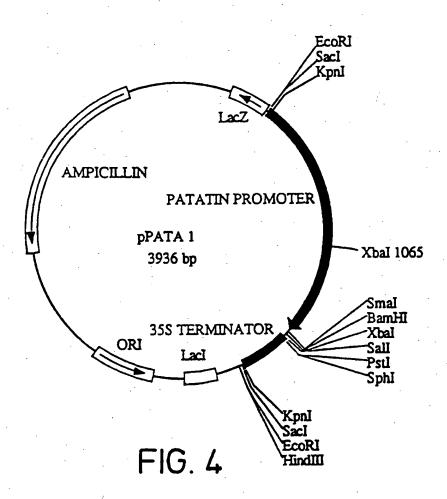


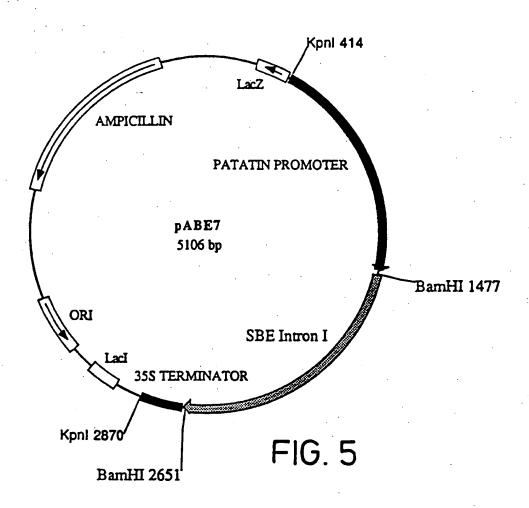
SUBSTITUTE SHEET (rule 26)

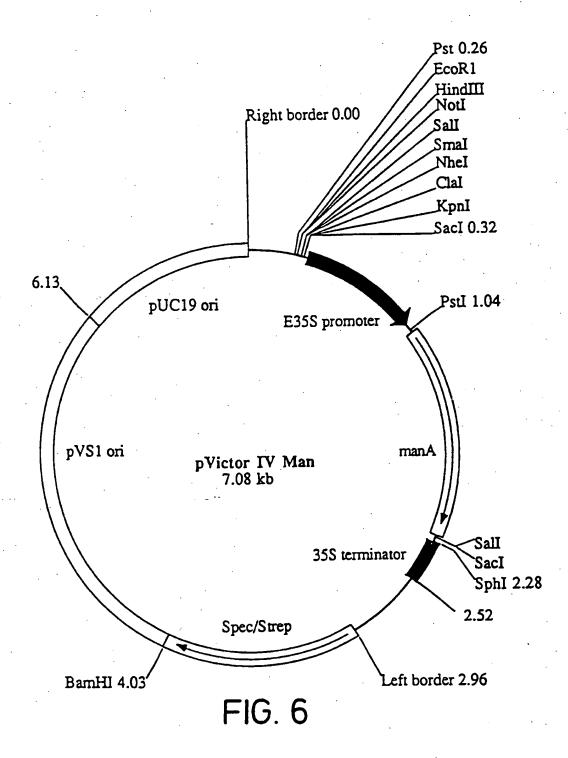




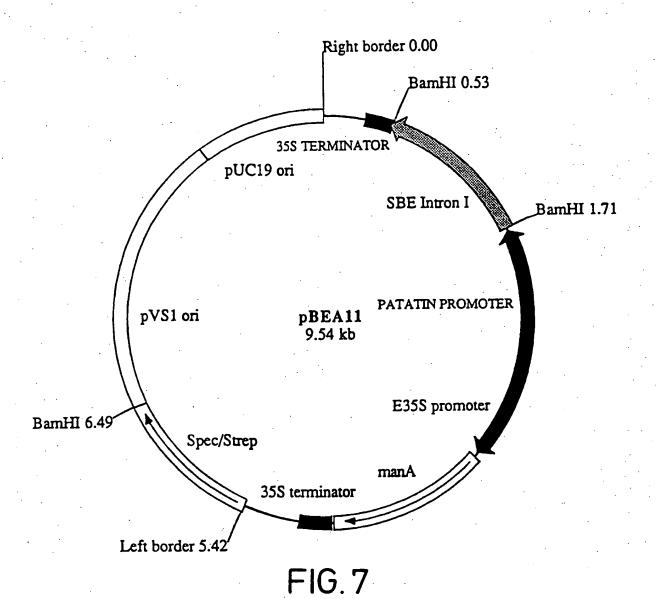
SUBSTITUTE SHEET (rule 26)







SUBSTITUTE SHEET (rule 26)



SUBSTITUTE SHEET (rule 26)

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GACCGGTCCTACT	PACAGACGATA(CTAACCCGTGG	AACTGTTGC	TCTGCTTCT	TAGAACT	120
CTATGGCTATTT	CGTTAGCTTG	GCGTCGGTTTC	BAACATAGTTT	TTGTTTCA	AACTCTT	180
CATTTACAGTCA	AAATGTTGTAT	GGTTTTTGTT	TCCTCAATG	ATGTTTACAG	TGTTGTG	240
TTGTCATCTGTA	CTTTTGCCTAT	TACTTGTTTTC	EAGTTACATG	rtaaaaaagt	GTTTATT	300
TTGCCATATTTT	GTTCTCTTATT.	ATTATTATCA:	PACATACATT	ATTACAAGGA	AAAGACA	360
AGTACACAGATC	TTAACGTTTAT	GTTCAATCAA(CTTTTGGAGG	CATTGACAGG	TACCACA	420
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CATAAAAATAGC	TAATGATAGAA	CATTGACATT	TGGCAGAGCT	TAGGGTATGG	TATATCC	540
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CGGTGGATATTA	TATTATGAGTT	GGCATCAGCA	AAATCATTGG	TGTAGTTGAC	TGTAGTT	660
GCAGATTTAATA	АТААААТССТА	ATTAACGGTC	Gatattaaaa	TAACTCTCAT	TTCAAGT	720
GGGATTAGAACT	'AGTTATTAAAA	AAATGTATAC	TTTAAGTGAT	TTGATGGCAT	TTTAATA	780
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ATTATAGTGTAA	ATTTATGCATI	CAGTGTAAAA	TTAAAGTATT	GAACTTGTCT	CTTTTAG	900
AAAATACTTTAT	ACTTTAATATA	AGGATTTTGTC	ATGCGAATTI) OTAATTAAK!	GATATTGA	960
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CTGAAAAATGCT	TGGCCATTCA(CAAAGTTTATO	TTAGTTCCT	ATGAACTTTA	TAAGAAGC	1260
TTTAATTTGAC	ATGTTATTTAT!	ATTAGATGAT!	ATAATCCATG	ACCCAATAGA	CAAGTGTA	1320
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AAAATAAATTA	ITTTTTGACAT	TCTAAAACTT	TAAGCAGAAT	AAATAGTTTA	TCAATTAT	1440
TAAAAACAAAA	AACGACTTATT	TATAAATCAA	CAAACAATTT	TAGATTGCTC	CAACATAT	1500

FIG.8

123456	10	20	30 45678901234	40 56789012	50 34567890		
TTTTCC	AATTAA	ATGCAGAAAA'	IGCATAATTTT	ATACTTG2	TCTTTATA	GCTTATTTTT	1560
TTTAGC	TAACC	ACGAATATTT	GTAAACTCACA	ACTTGATT	AAAAGGGA	TTTACAACAA	1620
GATATA'	rataagi	TAGTGACAAAT	CTTGÀTTTTAA	ATATTTT	LATTTGGAG	GTCAAAATTT	1680
TACCAT	AATCAT	ITGTATTTATA	ATTTAAATTTA	AATATCT	OATATITAT	ATATCTAGTA	1740
AACTTT	'ATAAAT	racgtatatac	AAAATATAAAA	TTATTGG	CCTTCATAT	TAGGTCAATA	1800
AATCCT	TAACTA'	TATCTGCCTTA	CCACTAGGAG?	AAGTAAA	AAACTCTTT	ACCAAAAATA	1860
CATGTA	TTATGT	ATACAAAAAGT	CGATTAGATTA	ACCTAAAT.	AGAAATTGT	TATAACGAGTA	1920
AGTAAG	TAGAAA'	OAAAAAATAT	TACAATACTA	TAĄAAAA	atgtttta(CTTCAATTTCG	1980
AAACTA	ATGGGG	TCTGAGTGAAA	TATTCAGAAA	GGGGAGGA	CTAACAAA	AGGGTCATAAT	2040
GTTTTT	ASTAIL	AAAGCCACTAA	LAATGAGGAAA?	ICAAGAAT	C <u>A</u> GAACATI	ACAAGAAGGCA	2100
GCAGCT	GAAGCA	AAGTACCATAI	TTTAATCAAT M		ATTTCAAA	GTTTTATCAAA V L S K	2160
ACCCAT	TCGAGG	ATCTTTTCCAT	CTTTCTCACC	TAAAGTTI	CTTCAGGG	•	2220
taattt	catgtt	aatttcaatt	atttttagcct	ttgcattt	cattttcc	aatatatctgg	2280
atcato	tcctta	gttttttatt	tatttttat	aatatcaa	atatggaa	gaaaaatgaca	2340
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gcctt	ctttata	attcgcgttgc	tttgaaatgct	tttactt	Lagccgagg	gtctattagaa	2760
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caaat	acatgg	tcaagggcaaa	gttctgaacad	cataaagg	gttcattal	catgtccaggga	2940
tatga	taaaaa	ttgtttctttg	rtgaaagttata	ataagatt	tgttatgg	cttttgctggaa	3000

FIG. 8 CONTINUED

SUBSTITUTE SHEET (rule 26)

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A S R N K I C F P S Q H S T G ACTGAAGTTTGGATCTCAGGAACGGTCTTGGGATATTTCTTCCACCCCAAAATCAAGAGT	3480
L K F G S Q E R S W D I S S T P K S R V TAGAAAAGATGAAAGGgtatgtttgataatttatatggttgcatggatagtatataaata	3540
R K D E R gttggaaaacttctggactggtgctcatggcatatttgatctgtgcaccgtgtggagatg	3600
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A V L T D D D N S T M A P L E E D V K T TGAAAATATTGGCCTCCTAAATTTGGATCCAACTTTGGAACCTTATCTAGATCACTTCAG	3960
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N R E D	G C I V Y aaatagtagtttccat	R E W A catcataacag	, P A A Q attttcctattaaagcatgatg	4680
			attaaggttatgcttctaatta	
		GTTATTGGCGA	ATTTCAATGGATGGAACGGTTCT	
	EAE	VIGD	F N G W N G S CTATTAGAATTCCTGATGTTGAC	
м и н и	EKDOF	GVWS	I R I P D V D MCGTTTCAAACATGGTAATGGA	
C K-P V	IPHNS	RVKF	R F K H G N G CCACTGCAGACGCCACAAAGTTI	
VWVD	RIPAW	IKYA	T A D A T K F CTTCAGAAAGgttttgttattca	
AAPY	DGVYW	D P P P	SER	
•			catttcgattcatgttcttacta	
•		*	aaataattgtgtctttactaati	
	·		gtcaattctataagtgcttgag	
		Y H F K		Τ.
GAGCCCCACG	AATCTATGAAGCACAT	TCGGCATGAG	CAGCTCTGAGCCACGTGTAAAT	r 5340 S
CGTATCGTGA	GTTTGCAGATGATGTT	TACCTCGGAT	TAAGGCAAATAACTATAATACT KANNYNT	G 5400 V
TCCAGTTGAT	GCCATAATGGAACAT	TCTTACTATGG	ATCATTTGGATATCATGTTACA	A 5460 N
	TGTGAGCAGTAGATAT		GGACCTAAAGTATCTGATAGAT	A 5520
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A H S ATGTCACTGA	L G L Q V TGGCCTCAATGGCTTT	L V D V GATATTGGCCA	V H S H A S N AGGTTCTCAAGAATCCTACTTI	N C 5640
V T D	GLNGF	DIGQ	G S Q E S Y F CAGGCTGTTCAACTATGCCAAT	H
AGE	RGYHK	L W D S	R L F N Y A N GGTGGCTAGAAGAGTATAACTT	W
EVL	RFLLS	NLRW	WLEEYNF	ָ ע
ĠFR	FDGIT	S M L Y	ATGTTCATCATGGAATCAATATC	G
FTG	NYNEY	FSEA	TACAGATGTTGATGCTGTGGT T D V D A V V	Y
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•					
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				ttacacttattaccgcc	
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				ctttctttacaatgaa	_1
•				ccgtctttcctcccct	
gryaractaarty	jaaaryyttiad	accigatati	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		

·		
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A Y Y R V I AGTGAGCTACTACCA	DERM	S E T	GTGACGAG			9360
a m 1 1. D 1	r a n T	FES	- D E	KPKD	2 6	
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ATTTCTAGTGAGCTA	CTACCAACAG	CAATATTG	AGGAGAGT	GACGAGAAAC	TAAAGAT	9480
T C C F 1. '	г. ъта	NIE	ES	DEKL	K D	9540
TCGTTATCTACAAAT	ATCAGTAACA'	MGATCAGA	CTGTTGTA	GTTCTGTTG	E R	, , , ,
S L S T N GACAAGGAACTTAAA	ISNI CATTURE COM	משטעעניטער T. Ö. G	·			9600
n w F 1. K	ns ps	VSI	. 1 5	ν	~ ~	
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14 D D C D	ANVW	GEI)		*	9720
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GAAATGGAGGAGTGATAGTCTCGAATATTATTCACCTCTTTAGCATTACCCGGTCTGGCT	11040
TTAAGGAGTTACGTCTTTTACGCTCGCCAATTTCTTTTTTAGAATGGTTGGT	11100
TCGCGAGTTGTGGAAGGTTCAAGTTACTCGATTCGTGATTTTCAAGTATGAGTGGTGAGA	11160
GAGATTCGATATTTTCACGAGGTGTATTCGAGGTCTAGTAGAACGAAGGGTGTCACTAAT	11220
GAAAGTTTCAAGAGTTCATCATCATCTTCTTAGTAGATTTTCGCTTTCAAATGAGTAT	11280
GAAAATTCTTCCTCTTTTCTATTGATTTTCTTCATTGTTTTCTTCATTGTTGTGTTGTT	11340
ATTGAAAAGAAAGAAAATTTATAACAGAAAAAGATGTCAAAAAAAGGTAAAATGAAAAGA	11400
GTATCATATACTTAAAGAGTTGCGTAGAGATAAGTCAAAAGAAACAGAATTATAGTAATT	11460
TCAGCTAAGTTAGAATTC	11478

FIG. 8 CONTINUED

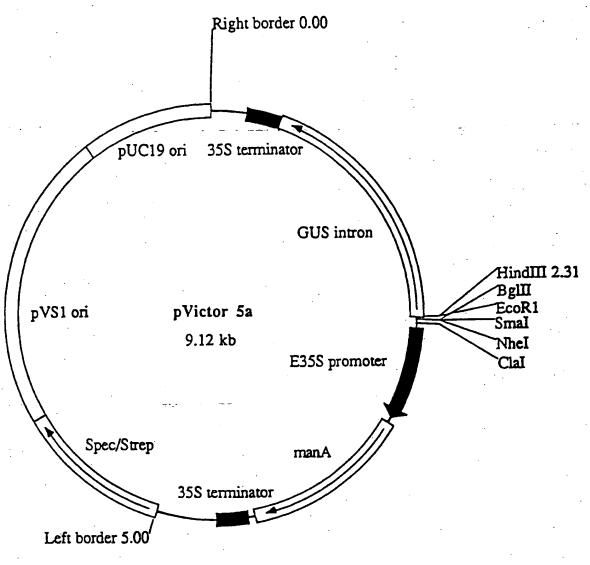
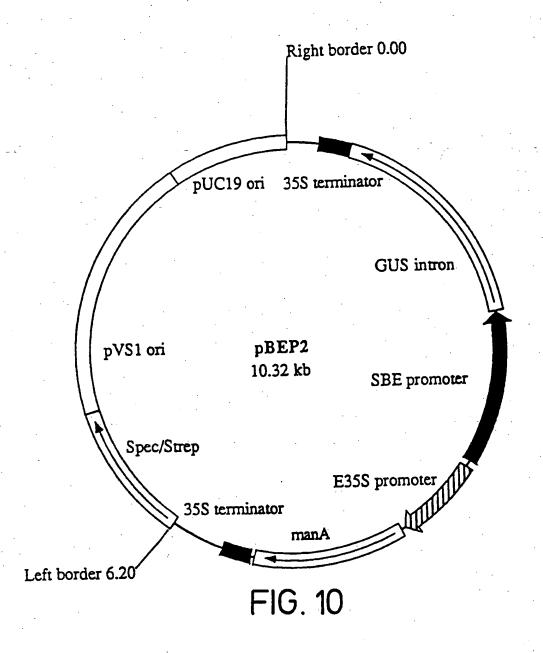


FIG. 9



SUBSTITUTE SHEET (rule 26)

18 / 25 KILAEKSSYNSESRPSTVAAS **ASRNKICFPSOHSTGLKFGSQ** INTRON 1: 1.2 kb INTRON 1: 2.0 kb MEINFKVLSKPIRGSFPSFSPKVSSG SBEII MYTLSGVRFPTVPSVYKSNGFSSNGDRRNANISVFLKKHSLSR EXON 1: 26 aa EXON 1: 44 aa SBEI

FIG. 11

SUBSTITUTE SHEET (rule 26)

10 20 30	40	50 60	
1234567890123456789012345678901	23456789012345	678901234567890	
	·		
GTATACACTCTCTGGAGTTCGTTTTCCTACT	TGTTCCATCAGTGTA V P S V Y	CAAATCTAATGGATT K S N G F	60
Bs			·
CAGCAGTAATGGTGATCGGAGGAATGCTAA' S S N G D R R N A N	TATTTCTGTATTCT	rgaaaaaacactctct k k h s l	120
BsaAI			
TTCACgtatgtctcactgtgtttgtggctg S R	tgtgtgttttttcl	cctgtctttttgtgtt	180
Bsp1286I BanII			240
ttgtgtaattggggctctttaaagttggta	ttgtgtataccctt	ttgagtatagtettig	240
aggaagcaaaatgatgaatcttgattgaca	ttagtaagggttgt	aactttttgaagtttg	300
gttaggtgtaattgagtttggcttgtgtgt	ctgtgtgtcgaggt	tatttttttggt tt gt	360
		· ·	
gttattggggatcttaaaagttggtattgt	gtatacccttttga	gtatagtetttgagg	a 420
	·		
agcaaaaatgatgaatcttgattggcatta	agtaaaggttgtago	ttttgaagtgtggt	± 480
aggtgtaattgagtttggcttgtgtgtctg	gtgtgttttggaato	cctgatgtgtgtcaag	t 540

FIG. 12

SUBSTITUTE SHEET (rule 26)

•						<u> </u>
10	20	30	40	50	60 67890	•
123456789012345678	19012345678	901234367	090123430	70701273	101020	
cctgatatgggtcgagg	tctttctttg	gtttgtgta	attgggggt	tcttaaaag	jttggt	600
	•					•
	•		ClaI			
			BspDI ▼			660
attatgtacctttttaa	gaatagtgtct	gagaaagca	aaatcgatg	aattttga	ttgaca	660
						•
				cttaatta	acatta	720
gcatattctttgagaaa	gcaaaaaatgg	gegagetete	catggagaaa	CLLyacty	acacca	, 20
	•					
ctaaaggtagcaacttt	ttcaactcct	gatatoggte	caaggttctt	tatttagt	ttgtgt	780
CLadayytaycaacccc	- CCCGGCCCCC	946463336				
	,					
			•			•
aatttggggttctttga	agttttgaga	aagaaaaat	tatgatttti	tcatggaga	aatttg	840
			Pvu	II		
AseI			Nspi	BII		
atttacattaataaagg	gtagtagcttt	ttaaagtgt	ggtcagctg	taatgagtt	cagctt	900
7.00	-1206T	•				•
	01286I 1II					
Apa	_	. *				960
ggtttaaaggggcccc	acatatggtg	ctttctggt	gagatattt	gttgctcc	accatac	960
		•	,			
	e e					
				catttttc	acttgac	1020
gagttataagaatcat	agegeeaggae				2000940	
•						
tagctactagaggagt	gatettgaege	rcggaaaato	cttagaaag	ggaaggtt	gtttgca	1080
	J=+g					
	_		_			

FIG. 12 CONTINUED

SUBSTITUTE SHEET (rule 26)

10 20 30 40 50 60 12345678901234567890123456789012345678901234567890	
Esp3I BsaBI	
tcaactggtgttatatgtgcaaggagacgggagatgatgtagatcatcttcttcatt	1140
gtggtctttccatgaggttatgatgtgatatgtttgaatggtttggtacttcttggctat	1200
EarI	
gccaagaactgtgaaagaattgatattcagttggaagtgtggaagttggaagagtggaaga	1260
attgacacttggttccattagctttaatgtgggtggtgtggagagaga	1320
ECORV agcttttgagggggtagagttgagctttcctcagttgagaagtagcctttgatatctttt	1380
EcoRI MumI ttttttttttttttgtacacccatagaattcccaattgtatagaagattgggtggagtttgt	1440
agagaatcatcttttgtagtagattctttaccttttggtatatccattgtatacagccag	1500
StuI gcctttgactatgtttatgaatgaatatacattacttgaaaaaaaa	1560
tctgttgtacctttgtagacaatgttgttgcagcatcttgataattccctgaaaattgtc	: 1620
FIG 12 CONTINUED	

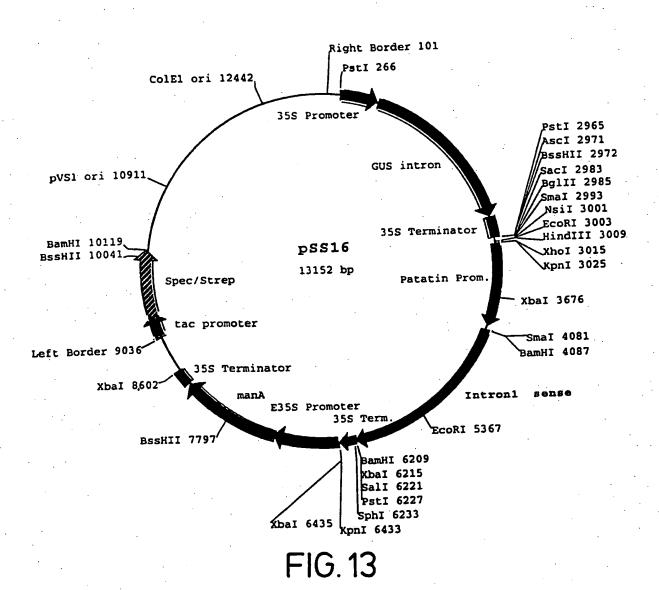
FIG. 12 CONTINUED SUBSTITUTE SHEET (rule 26)

•				
10 20	30	40	50 60 5678901234567890	· .
12345678901234567890	12345678901234	<u> </u>	0070301234707070	
•	-			
•			•	
tccctgaaggaatagtttgg	;ttgatattgattat	ttcttggttt	gtttaattcggtgttc	1680
· ·		•		
				1740
ttgaaggccattttaaatco	ctttgacattgttaa	aggtgtttac	aagtgttggtctgggt	- 1740
		•		
N			cctccaaaaaaaaaaat	1800
ttaaaagcacctcttgtat	ggtgctttctggagt	gattttttt	CCCCaaaagagaaga	
•	•			•
·		BclI Bgl	LII	
tgcaagaatcagtgtgtgt	acttttttctctto			1860
CACBAAGAAAAA	·	, ,		
cgttttagttgatttatcc	atatagtgaaagtt	ggtgtcatag	ttgctgtttgtggactt	1920
cctgtaaaagttttttgat	atacttaaaaaatt	gtcacacaga	agaaagagtttttacc	1980
			•	
			•	
AflII				2040
attacttaagctagatggg	jactgtttgattett	agaccaaaca	atgaacccccgcccc	
	•		•	
AflIII			•	٠
cttaacgtgtacttgaaat	Fagtttggtaaaatt	ot nat aggaa	aaaaqataattcttgat	2100
Cicaacycycacccyaaa				
			· ·	
	•		EarI	•
tgcttttggagcatcact	tctaatcataaaagt	ctttgctctc	ttcaaccatgaatgata	2160
		•		
		•		

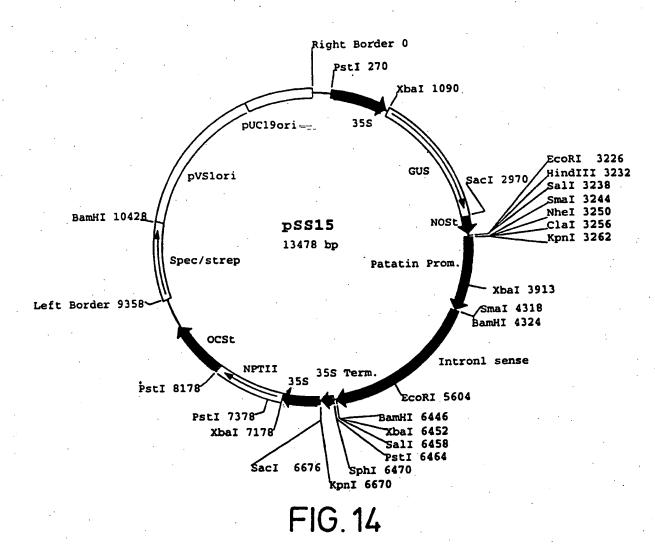
FIG. 12 CONTINUED SUBSTITUTE SHEET (rule 26)

10	20	30	40	50	60	
123456789012345	6789012345	6789012345	678901234	<u>5678901234:</u>	67890	
			,			•
aattggacacttatg	gtggccctaag	gttgctctcag	gtagtggtct	ttaattgtgga	agatat	2220
	•			•		٠
aactaatctgatata	atgtatgtag	BglII GGAAGATCTTV K I L	BDSI GCTGAAAAC A E K	TCTTCTTACA S S Y N	ATTCCG S E	2280
					•	•
s	fcI					. 22.00
AATCCCGACCTTCT	ACAGTTGCAG I V A A	CATCG S	•			2309

FIG. 12 CONTINUED



SUBSTITUTE SHEET (rule 26)



SUBSTITUTE SHEET (rule 26)

INTERNATIONAL. SEARCH REPORT

anal Application No PCT/TR 98/00295

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	SEARCHED ocumentation searched (classification system followed by classification)	tion cumbale)	
IPC 6	C12N C08B	·	
Documenta	tion searched other than minimum documentation to the extent that	such documents are included in the fields see	arched
		· ·	
Electronic o	data base consulted during the international search (name of data b	base and, where practical, search terms used	
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	cited in the application see the whole document		
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	see page 5, paragraph 3 - paragr see page 9, paragraph 2 - page 1 paragraph 1		
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